

ASSOCIATION OF miR-223-3p AND miR-223-5p GENE EXPRESSION WITH LEVELS OF IgE, VITAMIN D AND Mg²⁺ IN PEDIATRIC ASTHMA PATIENTSPOVEZANOST EKSPRESIJE GENA MIR-223-3P I MIR-223-5P SA NIVOOM IgE, VITAMINOM D I Mg²⁺ KOD PEDIJATRIJSKIH PACIJENATA SA ASTMOMDuaa F. Al-Mashhadani¹, Basima Q. Alsaadi^{2*}¹Ministry of Health, Baghdad, Iraq²Institute of Genetic Engineering and Biotechnology for Post-graduate Studies, University of Baghdad, Baghdad, Iraq**Summary**

Background: Asthma, a global non-communicable disease, significantly impacts public health. Severe cases cause high morbidity and mortality rates, with childhood asthma rates in Iraq reaching 20%. The study investigated the expression levels of *miR-223-3p* and *miR-223-5p* genes linked to pediatric asthma and the impact of IgE, Vitamin D, and Mg²⁺ on asthma severity.

Methods: A study of 140 children aged 1–10 years with allergic asthma was conducted in Baghdad from November 2023 to February 2024. The patients were divided into three groups: those under 1, those aged 1–5, and those over 5. Serum IgE, Vit D3, and Mg²⁺ levels were determined using the Immunoglobulin E Test Kit.

Results: The results indicated a significant increase in the level of IgE in patients (353.812±25.679 ng/mL) compared to the control group (25.320±2.581 ng/mL) (*p*-value <0.01, the serum level of Vit-D revealed a significant decrease in the patient group (16.907±0.512 ng/mL) while in the control group, levels reached (32.746±1.629 ng/mL), Mg²⁺ serum level decreased in patient groups (2.168±0.030 ng/dL) compared to control group (2.316±0.028 ng/dL) at *p*-value < 0.01. also, this study shows that upregulation of gene expression in *miR-223-3p* and *miR-223-5p* genes are considered risk factors for allergic asthma.

Conclusions: The study found that asthma patients had high IgE levels, low vitamin D3 and magnesium levels, and high gene expression in *miR-223-3p* and *miR-223-5p* genes, risk factors for allergic asthma.

Keywords: pediatric asthma, *miR-223-3p*, *miR-223-5p*, IgE, vitamin D, Mg²⁺

Kratak sadržaj

Uvod: Astma je globalna nezarazna bolest koja značajno utiče na javno zdravlje. Teški oblici astme uzrokuju visoku stopu morbiditeta i mortaliteta, pri čemu stopa dečje astme u Iraku dostiže 20%. Ova studija istraživala je nivoe ekspresije gena *miR-223-3p* i *miR-223-5p* povezanih sa dečjom astmom, kao i uticaj IgE, vitamina D i Mg²⁺ na težinu astme.

Metode: Studija je obuhvatila 140 dece uzrasta od 1 do 10 godina sa alergijskom astmom, a sprovedena je u Bagdadu od novembra 2023. do februara 2024. godine. Pacijenti su podeljeni u tri grupe: mlađi od 1 godine, uzrasta od 1 do 5 godina i stariji od 5 godina. Nivoi IgE, vitamina D3 i Mg²⁺ u serumu određeni su korišćenjem kompleta za testiranje imunoglobulina E.

Rezultati: Rezultati su pokazali značajno povećanje nivoa IgE kod pacijenata (353,812±25,679 ng/mL) u poređenju sa kontrolnom grupom (25,320±2,581 ng/mL) (*p*<0,01). Nivo vitamina D u serumu je bio značajno snižen kod pacijenata (16,907±0,512 ng/mL) u poređenju sa kontrolnom grupom (32,746±1,629 ng/mL). Nivo Mg²⁺ u serumu bio je niži kod pacijenata (2,168±0,030 ng/dL) u odnosu na kontrolnu grupu (2,316±0,028 ng/dL) uz *p*<0,01. Takođe, studija je pokazala da je povećana ekspresija gena *miR-223-3p* i *miR-223-5p* povezana sa povećanim rizikom od razvoja alergijske astme.

Zaključak: Studija je utvrdila da pacijenti sa astmom imaju visoke nivoe IgE, niske nivoe vitamina D3 i magnezijuma, kao i povećanu ekspresiju gena *miR-223-3p* i *miR-223-5p*, što predstavlja faktore rizika za razvoj alergijske astme.

Ključne reči: dečja astma, *miR-223-3p*, *miR-223-5p*, IgE, vitamin D, Mg²⁺

Introduction

Allergic asthma is a prevalent chronic inflammatory airway disorder characterised by various clinical manifestations and intricate genetic and causative variables (1, 2).

Allergic asthma is represented by airway inflammation, hyper-responsiveness, increased immunoglobulin (Ig) E levels, airway remodelling, and clinical manifestations including wheezing, dyspnea, chest tightness, cough, and airflow obstruction (3). In 2020, the World Health Organization (WHO) estimated that approximately 339 million individuals had asthma, with the majority of fatalities occurring among older adults. According to the Global Initiative for Asthma (GINA), asthma affects 1% to 18% of populations across various countries, and its prevalence has risen globally (3, 4).

Immunoglobulin E (IgE) is pivotal in the pathogenesis of asthma and other allergic conditions, including urticaria and allergic rhinitis. One idea regarding the rising incidence of asthma pertains to vitamin D (VitD). Specific individuals contend that many variables linked to westernisation have contributed to diminished (VitD) levels, leading to increased asthma prevalence. Conversely, some contend that Vitamin D exerts a more detrimental influence on allergy aetiology (5, 6). Magnesium (Mg⁺²) is a cofactor regulating several enzymatic and cellular activities. Another advantageous impact in asthma is reduced acetylcholine release from cholinergic neurons and diminished histamine release from mast cells. Enhanced bronchial smooth muscle contractility leading to bronchial hyper-reactivity is a defining pathophysiological event of asthma (7). Magnesium is a crucial factor in the contraction and relaxation condition of the bronchial smooth muscle. Magnesium positively influences bronchial asthma through various mechanisms, including direct bronchodilation and functioning as a natural calcium antagonist (8). Research indicates that Mg⁺² treats severe asthma (9, 10). Research on serum Mg⁺² concentrations in asthma patients indicates that hypomagnesemia is prevalent in individuals with asthma (11). MicroRNAs (miRNAs) are valuable diagnostic, prognostic, and therapeutic biomarkers for asthma. miRNAs are endogenous noncoding RNAs, approximately 19 to 25 nucleotides in length, involved in post-transcriptional gene regulation. Numerous miRNAs have been identified as contributors to lung formation, immunological responses, and many pulmonary illnesses, including lung cancer, asthma, COPD, and pulmonary fibrosis (12). Prior research has demonstrated that various miRNAs, such as miR-223-3p and miR-223-5p, exhibit upregulation or downregulation in individuals with asthma and correlate with increased adverse events, significant symptom burden, or accelerated decline in lung function (13). Various cell types generate miRNAs,

which are released into serum/plasma, saliva, bronchoalveolar lavage fluid (BALF), and other bodily fluids. Moreover, miRNAs are consistently found in circulating blood and other bodily fluids, where they perform biological functions via fluid circulation and serve as non-invasive biomarkers for diagnosing many disorders, including asthma. Recently, increased focus has been directed towards multifunctional miR-223 because of its crucial involvement in the immune system (14). The correlation between miR-223-3p expression in the peripheral blood leukocytes of asthmatic patients and inflammatory cytokines is not yet elucidated (15). There are many studies in Iraq about the relationship between genetics and asthma such as (4, 16–22). The aim of the current study is to investigate the relationship between gene expression and also the levels of Vitamin D3, IgE, and Mg⁺² with severity of asthma.

Materials and Methods

Subject

The current study was conducted on 140 subjects, 60 control and 80 patients (52 females and 88 males) with 1–10 years of allergic asthma. The patients were separated into three groups: The first group consisted of those under 1 year old, the Second group included those aged 1–5 years, and the third group included those beyond 5 years old, who were admitted to the Central Children's Hospital, Al-Kadhimiya Children's Hospital, Al-Alawiya Children's Hospital, and Al-Zahraa Center for Asthma and Allergy in Baghdad over the period from November 2023 to February 2024.

Blood sample collection

Five mL of venous blood was collected using a sterile syringe. Divide this blood into 3 mL, place it in a gel tube, and leave it for 20 minutes to clot at room temperature (25–30 °C). The tubes were then centrifuged at 3000 RPM over 15 minutes to separate the serum, and the serum was then stored in Eppendorf tubes at -20 °C until used for immunohistochemical assay. The remaining 2 mL of blood was placed in an EDTA tube. The remaining 2 mL of blood was placed in an EDTA tube. Then, 250 µL of blood sample was added to 750 µL in Trizol Eppendorf and frozen at -20 °C for molecular analysis.

Determination of vitamin D3

A solitary assessment of VitD, quantified as 25-hydroxy cholecalciferol, 25(OH)D, was conducted in all participants utilising a chemiluminescent technique (Liaison 25-OH Vitamin D Total; Diasorin, Saluggia, Italy). Values were treated as continuous variables, whereas vitamin D was classified in descrip-

tive analyses as desirable (or acceptable) at levels of at least 30 to 40 ng/mL (75 to 100 nmol/L), insufficient between 20 and 30 ng/mL (50 and 75 nmol/L), and deficient when below 20 ng/mL.

Determination levels of [Mg] in serum

Magnesium concentrations in biological samples were measured using the DRI-CHEM NX500i, an automated dry chemistry analyser developed by Fujifilm. The device is known for its high accuracy and ease of use, making it ideal for rapid and efficient analysis of clinical samples. The DRI-CHEM NX500i operates on the principle of dry chemistry, using reactive strips embedded with specific chemical reagents that interact with Mg^{+2} in the sample. The serum sample is introduced into the device, and the analysis is automatically performed. After processing the data, the device provides accurate results for Mg^{+2} concentration in the appropriate units.

Determination of serum IgE levels

Immunoglobulin E test kit (Rate Scattering Turbidimetric Method)

The IgE assay is used by modified dispersion turbidimetry to determine serum IgE levels to assess immune function and adjunctive diagnosis of immune diseases. IgE in the sample is bound to a specific antibody, which causes light to scatter and is proportional to IgE levels. A specialised device measures the intensity of the scattered light, and IgE concentrations are estimated by comparing the turbidity of the sample to a standard concentration. Results are recorded and repeated as needed, with control checks performed at the beginning of each batch to ensure accuracy and comparison of results to reference values.

Molecular study
Genomic RNA extraction

RNA was isolated from the blood of patients with asthma and a control group of apparently healthy individuals using the TransZol Up Plus RNA Kit (blood). The RNA content and purity were subsequently assessed using a Nanodrop spectrophotometer from the business Transgen.

cDNA synthesis for mRNA

The EasyScript® One-Step gDNA Removal and complementary DNA (cDNA) Synthesis SuperMix kit was used to reverse-transcribe total RNA into cDNA. Following the manufacturer’s guidelines, the reaction volume was set at 20 μ L, utilising 20 μ L of total RNA for the conversion process. as shown in Table I.

Table I Strand cDNA synthesis reaction component.

Component	volume reaction
mRNA/miRNA	4 μ L
Anchored Oligo(dT)18 Primer (0.5 μ g/ μ L)	1 μ L
Random Primer (0.1 μ g/ μ L)	1 μ L
GSP	1 μ L/10 pmol
2xES Reaction Mix	10 μ L
EasyScript ® RT/RI Enzyme Mix	1 μ L
gDNA Remover	1 μ L
RNase-free Water	1 μ L
Total volume	20 μ L

Table II Primer sequences were utilised in this study’s assays.

Primer	Sequence (5′→3′ direction)	primer size (bp)	T _a (°C)	Design in the current study
miRNA				
MiR-223-3P	TGTCAGTTTGTCAAATACCCCA	22	58	
MiR-223-5P	CGTGTATTTGACAAGCIGAGTT	22		
miRNA-universe R.P.	GCGAGCACAGAATTAATACGAC	22		
Universe R.transcription p	CAGGTCCAGTTTTTTTTTTTTTTTTTVN	26		
MiR -U6	AGAGAAGATTAGCATGGCCCCT	22	58	

*Ta: annealing temperature

Gene expression of MiR-223-3P and MiR-223-5P by quantitative real-time PCR (qRT-PCR)

Total RNA was reverse transcribed into complementary DNA (cDNA) with the EasyScript One-Step gDNA Removal and cDNA Synthesis SuperMix Kit from TransGen Biotech Co., China, in a reaction volume of 20 µL, adhering to the manufacturer's instructions. The quantitative real-time PCR (qRT-PCR) was performed utilising the QIAGEN Rotor-Gene Q real-time PCR system (Germany). Each qRT-PCR reaction involved 2 µL of cDNA, 1 µL for both the forward and reverse primers (with a concentration of 10 µmol/L) as listed in *Table II*, and 10 µL of the PerfectStart™. The green qPCR SuperMix kit from TransGen Biotech Co., China. The thermal profile consisted of an initial step at 94 °C for 5 minutes (one cycle), followed by 40 cycles involving denaturation at 94 °C for 5 minutes, annealing at 58 °C for MiR-223-3P, MiR-223-5P, and miR-U6 for 15 seconds, and extension at 72 °C for 20 seconds. The final dissociation stage spanned from 55 to 95 °C, with each degree lasting 5 seconds. The specificity of the amplified product was confirmed through melting curve analyses. The relative expression of the MiR-223-3P and MiR-223-5P genes in the study group samples was evaluated by normalising their expression levels to the reference gene miR-U6 using the Ct1 method (20). The data were presented as the fold change in MiR-223-3P and MiR-223-5P gene expression compared to the healthy controls within the study groups. This allowed for normalising the expression levels against the reference gene miR-U6. The median fold expression of MiR-223-3P and MiR-223-5P in the study groups was then utilised to assess whether there were statistically significant differences in MiR-223-3P and MiR-223-5P gene mRNA expression levels. Significant differences.

MiR-223-3P and MiR-223-5P gene expression calculation

The fold changes in the quantitative expression of mature RNAs were assessed using the relative cycle threshold (2- $\Delta\Delta C_t$) method, initially introduced by Livak and Schmittgen in 2001. It is the ratio of relative gene expression between the control group and the experimental group. The double delta Ct (threshold cycle) analysis was used to assess the expression of MiR-223-3P and MiR-223-5P genes, the housekeeping reference genes. The calculations were as follows: The real-time cyclor software calculated the threshold cycle (CT) for each sample. The samples were duplicated, and the average results were computed. The Ct values for the target genes MiR-223-3P and MiR-223-5P, which were being evaluated in both patients and controls, were reported.

The ΔC_t , or difference in CT values, also called the »normalised raw data,« was determined by sub-

tracting the specified normalisation factor from the Ct value of each target gene and the housekeeping gene.

$$\Delta C_t (\text{control}) = C_t (\text{gene}) - C_t (\text{HKG})$$

$$\Delta C_t (\text{patient}) = C_t (\text{gene}) - C_t (\text{HKG}) \quad \Delta\Delta C_t = \Delta C_t (\text{patient}) - \Delta C_t (\text{control}).$$

Ethical approval

The study used the ethical principles outlined in the Declaration of Helsinki. The study was performed following the acquisition of both verbal and written consent from the patients before collecting the samples. This case-control study was approved by the Scientific Committee of the Institute of Genetic Engineering and Biotechnology for past graduate study at the University of Baghdad, and the study was approved by the Ministry of Health and Environment of Iraq (4778 on 25-12-2023).

Statistical analysis

The results of the present study were analysed related to the objectives and presented according to the general description of the sample. Microsoft Excel 2010 and SPSS (version 25) software were used for statistics analysis. Microsoft package (Excel and Word). The data are expressed as mean \pm SD, and differences were considered significant when p-values were $P < 0.05$.

Results

Serum IgE, vitamin D and Mg⁺² levels in studied groups

The current study showed the levels of IgE in the blood serum of both patients and control groups. The results indicated a significant increase in the level of IgE in patients (353.812 ± 25.679 ng/mL) compared to the control group (25.320 ± 2.581 ng/mL) (p-value < 0.01) as shown in *Figure 1*. The serum level of Vit-D revealed a significant decrease in the patient group (16.907 ± 0.512 ng/mL), while in the control group, levels reached (32.746 ± 1.629 ng/mL), as shown in *Figure 2*.

The miR-223-3p and miR-223-5p gene expression

The results demonstrated that using miR-U6 for normalisation in qRT-PCR is a highly reliable approach, particularly in clinical studies. Furthermore, the 2-ct value and the ratio of 2-ct for various study groups compared to the control group were employed to assess changes in miR-U6 expression across different study groups, as presented in *Table*

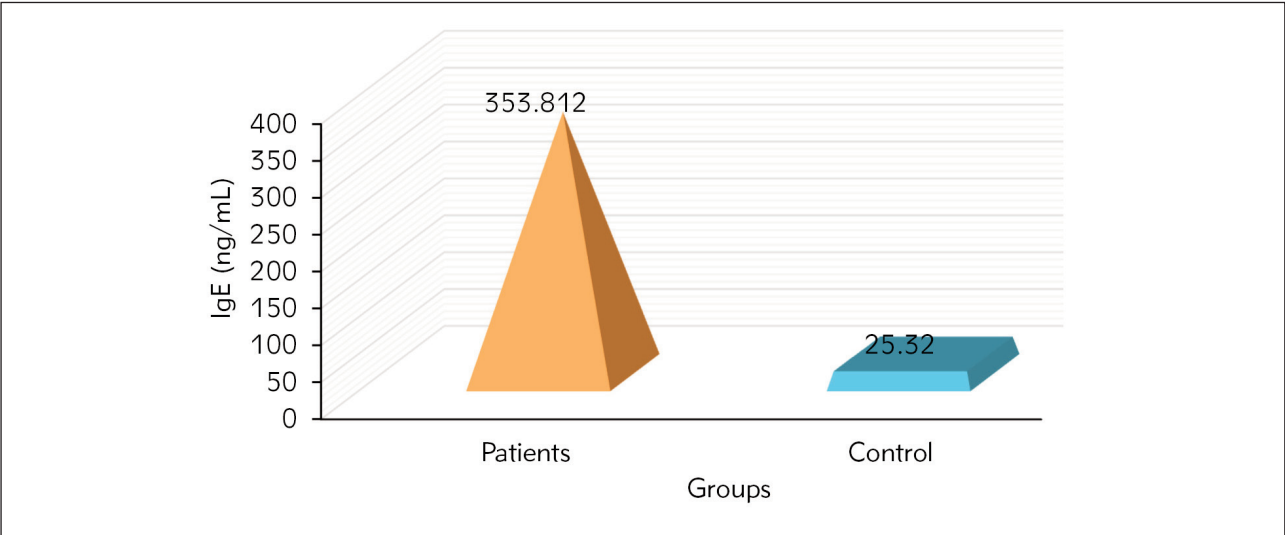


Figure 1 Comparison between the IgE(ng/mL) levels between patients and control groups.

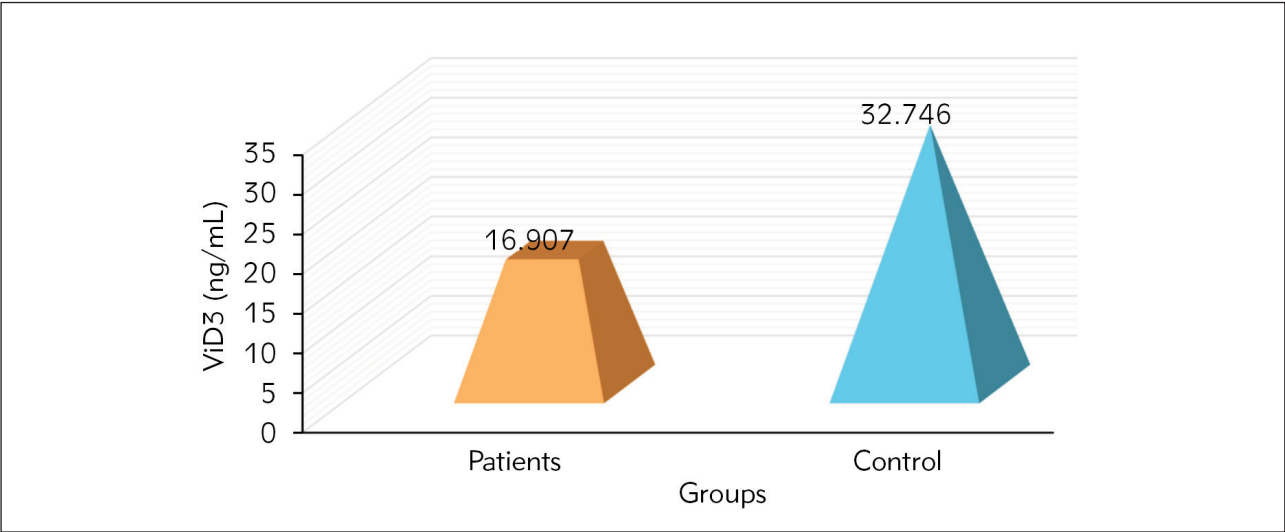


Figure 2 Comparison between the Vit.D3 between patients and control groups.

III. The $2^{-\Delta\Delta Ct}$ value for asthma patients was 304.7537; for control, it was 258.078. The calculated fold expression ratios for the gene were 1.180 and 1.00, respectively. These variations in gene fold expression among the study groups underscore the utility of the miR-U6 gene as a reliable control. The expression of the miR-223-3p gene showed highly significant differences ($p < 0.001$) in the asthma patient group compared to the control group. Similarly, Table IV shows the $2^{-\Delta\Delta Ct}$ value for asthma patients was 18.600, and for control, it was 8.141. The calculated fold expression ratios for the gene were 2.29 and 1.00, respectively. The expression of the miR-223-5p gene showed highly significant differences ($p < 0.001$) in the asthma patient group compared to the control group.

Quantification of miR-223-3p expression by real-time PCR

The qPCR samples of the studied groups determined miRNA-223-3p dissociation curves. Table III shows the results of miRNA-223-3p levels in children and patients infected with allergic asthma, and healthy controls are demonstrated in Table III. The folding of miRNA-223-3p (1.180) revealed higher levels in patients with allergic asthma and (1) in apparently healthy controls, as shown in Figures 3 and 4.

Table III Comparison between patients and healthy control groups regarding miR-223-3p fold expression levels.

Groups	Means Ct ofmiR223-3p	Means Ct ofU6	DCt (Means Ct of miR223-3p)	2 ^{-ΔCt}	experimental group/ Control group	Fold of gene expression
Patients	16.4761	24.7276	-8.2515	304.7537	304.7537/258.078	1.180
Control	16.2341	24.2458	-8.01166	258.078	258.078/258.078	1.00

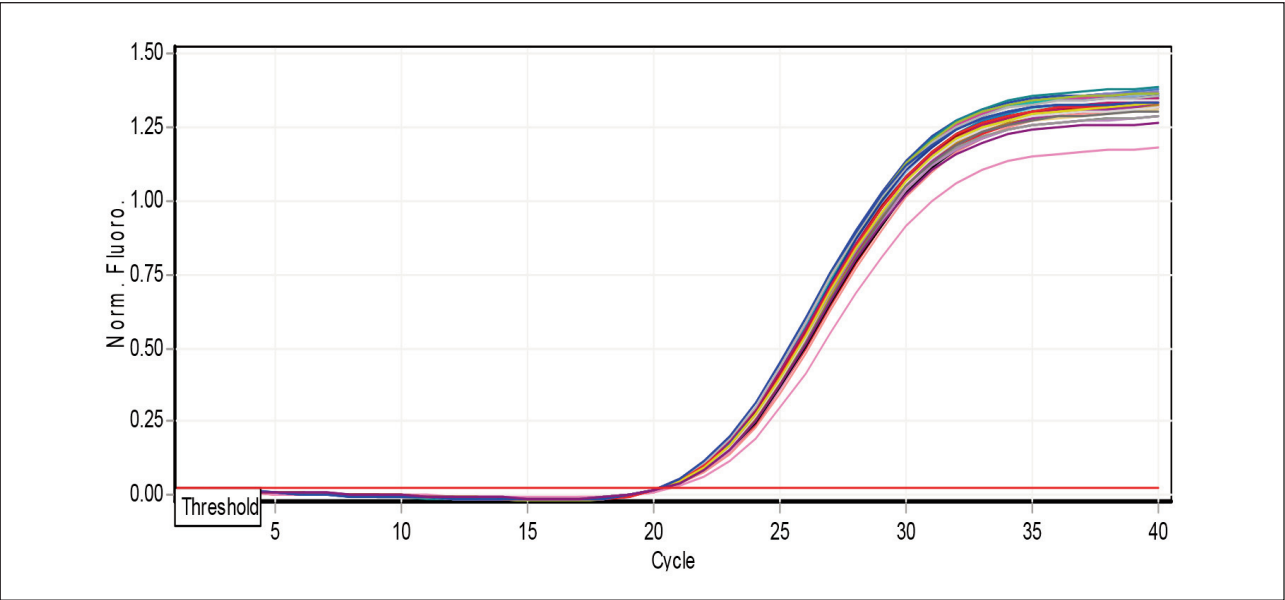


Figure 3 Amplification curve chart miR-223-5p gene.

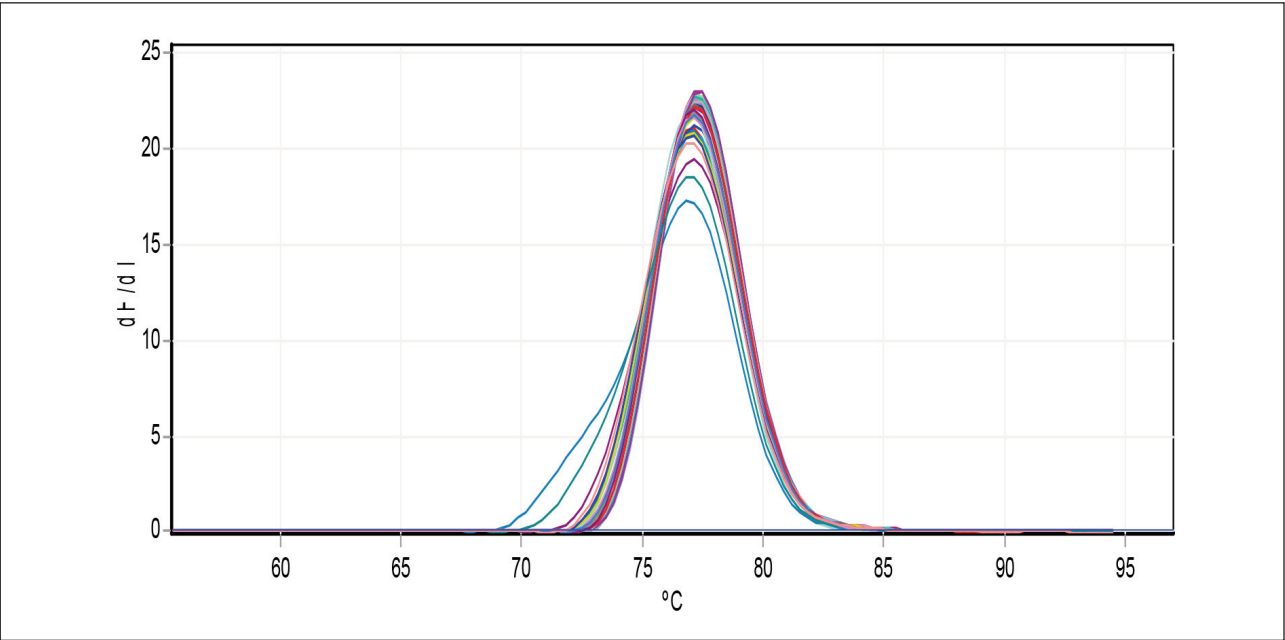


Figure 4 Melting curve chart miR-223-3p gene.

Table IV Comparison between patients and healthy control groups regarding miR-223-5p fold expression levels.

Groups	Means Ct ofmiR223-5p	Means Ct ofU6	Δ Ct (Means Ct of miR223-5p)	$2^{-\Delta Ct}$	experimental group/ Control group	Fold of gene expression
Patients	20.5103	24.7276	-4.21725	18.600	18.600/8.141	2.29
Control	21.2371	24.2625	-3.0252	8.141	8.141/8.141	1.00

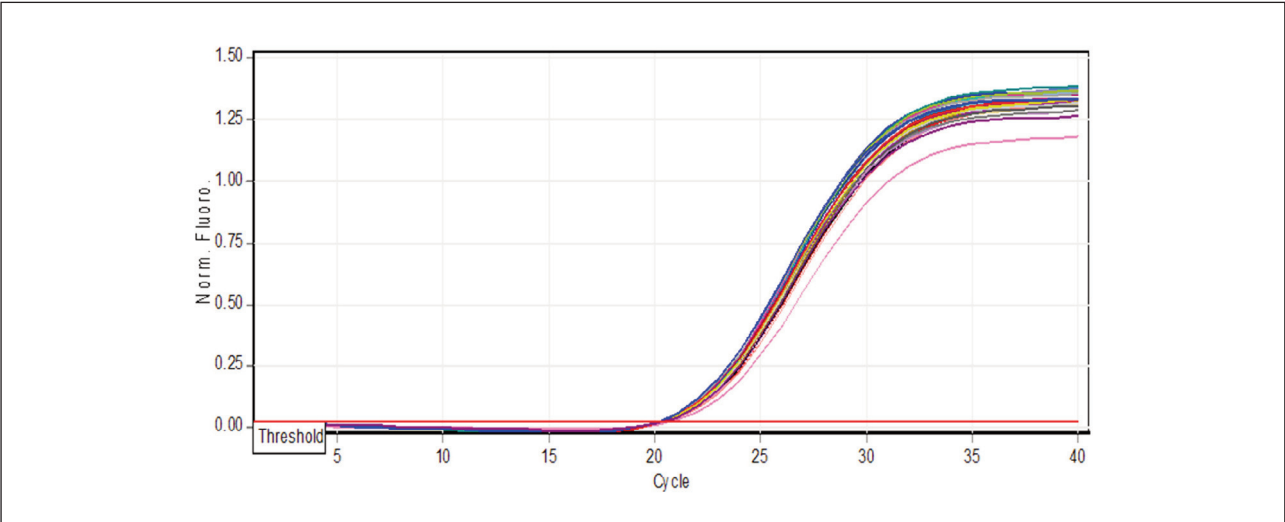


Figure 5 Amplification curve chart *miR-223-5p* gene.

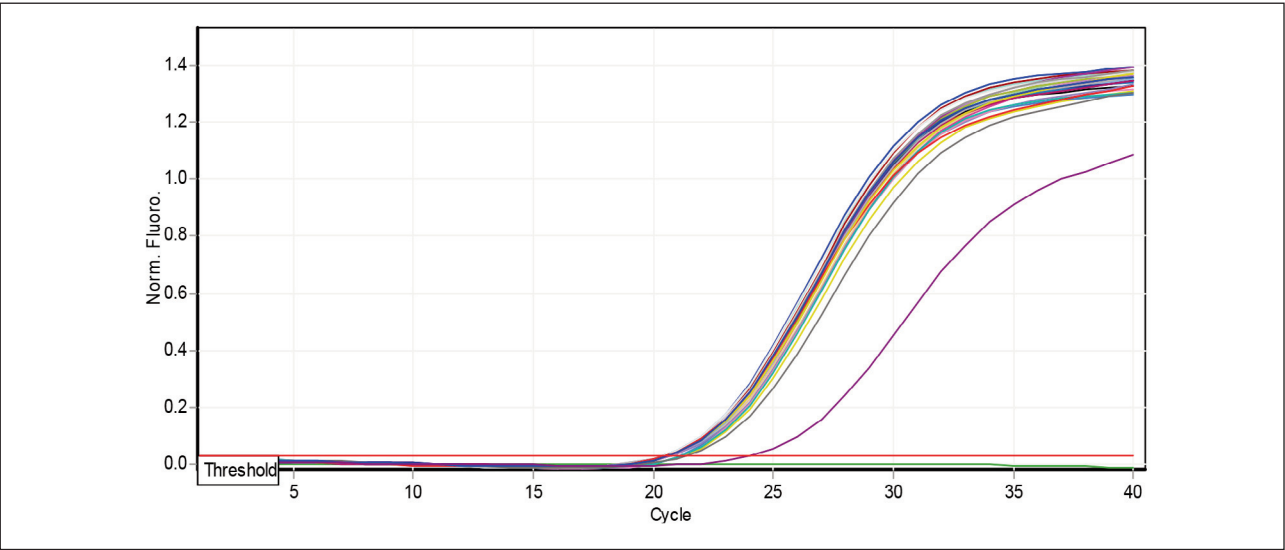


Figure 6 Melting curve chart *miR-223-5p* gene.

Quantification of miR-223-5p expression by real-time PCR

miRNA-223-5p dissociation curves by q PCR samples of studied groups. The results of miRNA-223-5p levels in children patients infected with aller-

gic asthma and healthy controls are demonstrated in Table IV. The folding of miRNA-223-5p (2.29) revealed higher levels in patients with allergic asthma and (1) in apparently healthy controls, as shown in Figures 5 and 6).

Table V Pearson correlations for miR-223-3p and miR-223-5p.

		miR223-3p	miR223-5p
miR223-3p	Pearson Correlation	1	.659**
	Sig. (2-tailed)		<.001
miR223-5p	Pearson Correlation		1
	Sig. (2-tailed)		

**Correlation is significant at the 0.001 level (2-tailed).*

In Table V, the Pearson correlation between miR-223-3p and miR-223-5p showed a strong positive correlation of 0.659 with a statistical significance of less than 0.001. This correlation suggests a possible co-regulatory relationship or involvement in a specific biological pathway, which explores their role in childhood asthma and its associated biological mechanisms.

Discussion

Immunoglobulin E (IgE) plays a crucial role in allergic diseases, particularly asthma, due to its heightened sensitivity to allergens; hence, measuring IgE levels aids in diagnosing asthma and monitoring patient status. The present study showed elevated total serum IgE of asthmatic compared to control at (P<0.001). The result agrees with previous studies (23). Another study by (24) indicates that the mean serum IgE level was 554 IU/mL in asthmatic patients, while that of the control was 69 IU/mL. Further, the present results agree with a previous international study, Strømgaard et al. (25) identified a positive correlation between total blood IgE levels and asthma in patients . Elevations in IgE levels may be attributed to viral factors (the predominant cause of asthma symptom aggravation), specific allergens, or may merely indicate a broad elevation of IgE synthesis. Consequently, IgE levels can be utilised to distinguish between asthmatic and non-asthmatic individuals (26). The current study showed a decrease in vitamin D in apathetic patients compared to control. These findings are consistent with the results obtained from other studies, both adults and children (27).

The active form of vitamin D can inhibit inflammation by enhancing the secretion of anti-inflammatory cytokines and chemokines. So, vitamin D’s endocrine, autocrine, paracrine, and immune-modulating activity emerged in the body (28). Another study demonstrated that incorporating VitD3 supplementation into inhaled corticosteroid therapy for asthma did not significantly impact the duration until severe asth-

ma developed or asthma-related morbidity (29). Furthermore, VitD supplementation did not markedly decrease the incidence of initial treatment failure. Additionally, no observable effects on asthma control, pulmonary function, asthma symptoms, quality of life, or airway inflammation were noted (30).

In this study, the serum Mg⁺² level of asthmatic children was significantly lower compared to the control group. In agreement with (31) Lytvynets LI. (32) that there was no significant difference in serum levels between asthmatics and controls; nevertheless, they discovered that asthmatics had lower levels of intracellular magnesium in their erythrocytes. His discovery may offer medicinal advantages. Magnesium therapy for asthmatic patients may enhance clinical outcomes, as indicated by (33). Low Mg⁺² levels may, according to a study by Jebur and Saud (6), be associated with worsening asthma symptoms, with patients having lower levels than healthy controls. These findings are consistent with the current study, as evidence suggests that Mg⁺² deficiency may play a role in worsening asthma symptoms and is a potential factor influencing disease progression (8). It is possible that the clinical condition of asthmatic patients could be improved by using Mg⁺² supplements. Anti-inflammatory drugs, also known as glucocorticoids, and bronchodilator agents, also known as beta-2 agonists, are among the medications utilised to treat asthma. Patients who use these medications for an extended period may experience a decrease in magnesium levels due to urine excretion and intracellular shift (34).

MiR -U6 is one of the most commonly employed housekeeping genes for assessing gene expression data (35). In a study conducted by Robert and colleagues (36), the expression of 1,718 genes across 72 different types of normal human tissues was investigated using quantitative real-time polymerase chain reaction (qRT-PCR), with miR -U6 serving as a reference gene.

This study showed increased Mir-223-3p expression in severe asthma cases in children. This is consistent with the previous study’s findings (15), which found that the expression levels miR-223-3p in peripheral blood leukocytes were significantly higher in asthma patients than in healthy children. Some results have been reached by another study (2016), which indicated an increase in the expression of the Mir-223-3p, miR-629-3p, and miR142-3 genes, indicating the possibility of its contribution to the exact inflammatory mechanism related to Increased expression of mir223-3p is considered as a biomarker in the diagnosis of allergic asthma in children. A study conducted by Xu et al. (15) on the development of asthma through several genetic and molecular analyses clarified the role of mir-223-3p in developing and exacerbating the disease in childhood. Numerous studies have shown significant shifts in miR-223

expression during the development of inflammatory states. This predisposition points to plausible significant functions of this miRNA in maintaining a balanced inflammatory state (37). A study similar to the current study showed that the most critical biological study of miR-223-3p pertains to cilium construction and organisation in bronchial epithelial cells, facilitating mucociliary function and mucus clearance, hence reducing infections and inflammation (38). miR-223-3p is involved in the inflammatory response, and its expression level significantly changes in children with allergic asthma. Therefore, it is vital to explore the correlation between miR-223-3p and allergic asthma (39). Recent investigations into asthma have shown that exosomes play a part in the development of asthma. The results showed that the exosome derived from neutrophil swarm 1 contains miR-223-3p, which helps delay the inflammatory response in asthma (40). In addition, airway-derived miR-223-3p suppresses the expression of TNF- α -stimulated gene 6 in peripheral white blood cells, promoting asthma development (41). These results demonstrate that miR-223-3p plays a dual role in regulating the inflammatory response associated with asthma.

This study showed a significant increase in the expression levels of miR-223-5p, consistent with previous studies. As explained by earlier data (42), they conducted experiments on mouse models of asthma and revealed a similar increase in the expression of miR-223-5p in the affected models. MiR-223-5p is involved in various autoimmune disorders in children. Micro RNA223-5p may aim at an immunosuppressive effect to control inflammation or remain up-regulated to favour disease progression and susceptibility to infection MiR-223 (43) by driving the switch between effective regulatory cells of B and T cells and dendritic cells, may contribute to immune interaction and control immunomodulation (44). The importance of miR-223-5p in asthma pathogenesis and disease exacerbation has received significant attention in asthma (45). Recent research has shown that the miR-223 family of microRNAs, including Mir223-5p, which is highly selectively expressed in granulocytes,

is critical in controlling neutrophil function (46). The major function of miR-223-5p is to repress gene expression at the post-transcriptional level. In macrophages, statistical histogram analysis and analysis have identified possible miR-223-5p target genes involved in the inflammatory response, such as Ccl3, Tnf, Cxcl2, and Lcn2. A condition known as a microvascular complications-independent asthma predictor enhances miR-223-5p expression via the p38 (47). This shows that miR-223-5p expression is more closely linked with asthma than miR-223-3p. The conclusion reached was that miR-223 was involved in lung protection from chronic inflammation and played a crucial role in those mechanisms that developed within an inflammatory process, thereby in diseases such as asthma (38).

Conclusion

Current results indicate that serum the levels of IgE in the blood serum showed a significant in the patients compared to the control; the serum level of Vit-D revealed a considerable decrease in the patient group than in the control group, and magnesium levels are lower in asthmatics, and depending on the patients' magnesium condition, these variations were substantial. In addition, the *miR-223-3p* and *miR-223-5p* may be regarded as risk factors for the onset of asthma in the Iraqi population. To validate these results, larger sample sizes and additional research are required.

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Conflict of interest statement

All the authors declare that they have no conflict of interest in this work.

References

1. Ji Y, Wang E, Mohammed MT, Hameed N, Christodoulou M-I, Liu X, et al. Selective production of IL-33-neutralizing autoantibody ameliorates asthma responses and severity. *Clinical Immunology* 2024; 264: 110234.
2. Kadhem AA. The Role of IL-37 in Allergic Rhinitis, Asthma and Urticaria Diseases in Samples of Iraqi Patients. *Iraqi Journal of Science* 2024; 2418–30.
3. Atta R, Aloubaidy R. Genetic polymorphism of asthma in Iraq. *Iraqi Journal of Agricultural Sciences* 2022; 53(2): 288–96.
4. AL-Qadhi IY, AL-Saadi BQ. Impact of IL-4R (rs1805011) Gene Polymorphism on IL-4 Serum Level in Iraqi Allergic Asthma Patients. *Iraqi Journal of Biotechnology* 2022; 21(2).
5. Searing DA, Zhang Y, Murphy JR, Hauk PJ, Goleva E, Leung DY. Decreased serum vitamin D levels in children with asthma are associated with increased corticosteroid use. *Journal of Allergy and Clinical Immunology* 2010; 125(5): 995–1000.
6. Jebur MS, Saud AM. Serum levels of total IgE and interleukin-13 in a sample of allergic asthma patients in Baghdad. *Iraqi Journal of Science* 2020; 3208–14.

7. Ibrahim MS, Mohammed MM. Effect of pharmacist interventions on pulmonary function parameters of Iraqi asthmatic patients: A comparative study. *Iraqi Journal of Pharmaceutical Sciences* 2023; 32(2): 9–17.
8. Saud AM, Jebur MS. Strong Association of STIP1 Gene rs2236647 Polymorphism and Serum Magnesium Level with Bronchial Asthma in a Population from Iraq. *Journal of Bioscience and Applied Research* 2024; 10(3): 328–37.
9. Kaye P, O'Sullivan I. The role of magnesium in the emergency department. *Emergency Medicine Journal* 2002; 19(4): 288–91.
10. Cheuk D, Chau T, Lee S. A meta-analysis on intravenous magnesium sulphate for treating acute asthma. *Archives of Disease in Childhood* 2005; 90(1): 74–7.
11. Abuabat F, AlAlwan A, Masuadi E, Murad MH, Jahdali HA, Ferwana MS. The role of oral magnesium supplements for the management of stable bronchial asthma: a systematic review and meta-analysis. *NPJ Primary Care Respiratory Medicine* 2019; 29(1): 4.
12. Jiang Z, Zhang L, Shen J. MicroRNA: Potential biomarker and target of therapy in acute lung injury. *Human & Experimental Toxicology* 2020; 39(11): 1429–42.
13. Pattarayan D, Thimmulappa RK, Ravikumar V, Rajasekaran S. Diagnostic potential of extracellular microRNA in respiratory diseases. *Clinical Reviews in Allergy & Immunology* 2018; 54: 480–92.
14. Haneklaus M, Gerlic M, Kurowska-Stolarska M, Rainey A-A, Pich D, McInnes IB, et al. Cutting edge: miR-223 and EBV miR-BART15 regulate the NLRP3 inflammasome and IL-1 production. *The Journal of Immunology* 2012; 189(8): 3795–9.
15. Xu W, Wang Y, Wang C, Ma Y, He S, Kang Y, et al. Increased miR-223-3p in leukocytes positively correlated with IL-17A in plasma of asthmatic patients. *Iranian Journal of Allergy, Asthma and Immunology* 2020; 289–96.
16. Goodi GAK, AL-Saadi BQ. Polymorphism of FOXO3a Gene and Its Association with Incidence of Asthma in Iraqi Patients. *Iraqi Journal of Biotechnology* 2018; 17(3).
17. ALSaadi BQH, Ali MH. Association of serum level and gene expression of IL-25 in a sample of Iraqi asthmatic patients. *Revis Bionatura* 2023; 8(4): 84.
18. Abdulkareem ZT, AL-Saadi BQH. Determine gene expression of IL-17 in Iraqi Child Asthmatic Patients. *Iraqi Journal of Biotechnology* 2020; 3(19).
19. Goodi GAK, AL-Saadi BQ. The Relationship of Serum Levels of Tumor Necrosis Factor (TNF) Cytokine with Asthma. *Journal of Pharmaceutical Sciences and Research* 2019; 11(1): 206–7.
20. Faeqali Jan M, Muneer Al-Khafaji H, Hasan Al-Saadi B, Aneed Al-Saedi M. Assessment of interleukin-8 in bronchial asthma in Iraq. *Archives of Razi Institute* 2021; 76(4): 913–23.
21. Al-Saadi BQH, Al-Khafaji HM, Al-Saedi MKA. Primer and Probe Designing to Detect SNP rs 4073 in Interleukin-8 Gene in Iraqi Patients with Bronchial Asthma. *Journal of Applied Sciences and Nanotechnology* 2021; 1(3): 51–7.
22. AL-Qadhi, I.Y.; AL-Saadi, B.Q.H. Polymorphism of IL13 (rs1295685) Gene and Its Serum Level in a Sample of Iraqi Patients with Allergic Asthma. *Revista Bionatura* 2023; 8 (2) 63.
23. Hameed RM, Ahmed MM, Abood HAAN, Hussein AM. To evaluate total serum immunoglobulin E level and factors that effect on this level in Iraqi asthmatic children. *Biomedical and Biotechnology Research Journal (BBRJ)* 2019; 3(4): 240–4.
24. Ahmad Al Obaidi AH, Mohamed Al Samarai AG, Yahya Al Samarai AK, Al Janabi JM. The predictive value of IgE as biomarker in asthma. *Journal of asthma* 2008; 45(8): 654–63.
25. Strømgaard S, Thomsen SF, Fenger M, Backer V. Predictors of serum total IgE in a random sample of 7–17 year old children. *International Scholarly Research Notices* 2011; 2011(1): 169859.
26. Salman A-J, Wahab R. Association of Adiponectin and IgE levels with occurrence of asthma in Babylon Province. *Indian Journal of Forensic Medicine & Toxicology* 2020; 14(2).
27. Forno E, Bacharier LB, Phipatanakul W, Guilbert TW, Cabana MD, Ross K, et al. Effect of vitamin D3 supplementation on severe asthma exacerbations in children with asthma and low vitamin D levels: the VDKA randomised clinical trial. *Jama* 2020; 324(8): 752–60.
28. Wang Q, Ying Q, Zhu W, Chen J. Vitamin D and asthma occurrence in children: A systematic review and meta-analysis. *Journal of Pediatric Nursing* 2022; 62: e60–e8.
29. Li Q, Zhou Q, Zhang G, Tian X, Li Y, Wang Z, et al. Vitamin D supplementation and allergic diseases during childhood: a systematic review and meta-analysis. *Nutrients* 2022; 14(19): 3947.
30. Jat KR, Goel N, Gupta N, Gupta CP, Datta S, Lodha R, et al. Efficacy of vitamin D supplementation in asthmatic children with vitamin D deficiency: a randomised controlled trial (ESDAC trial). *Pediatric Allergy and Immunology* 2021; 32(3): 479–88.
31. Pirdawo DR, Alrababaty AA. The Relation between Serum Magnesium Level and Acute Exacerbation of Asthma in Children Admitted to Rapareen Teaching Hospital in Erbil City. *AMJ (Advanced Medical Journal)* 2018; 4(2): 53–7.
32. Lytvynets LI. Macro-and microelements imbalance in etiology and progression of bronchial asthma in children. *Likars'ka sprava* 2013(4): 33–8.
33. Hill J, Micklewright A, Lewis S, Britton J. Investigation of the effect of short-term change in dietary magnesium intake in asthma. *European Respiratory Journal* 1997; 10(10): 2225–9.
34. Kundu TK, Dutta A, Chatterjee A, Chowdhury A. Association between Serum Magnesium to Calcium Ratio with Level of Asthma Control in Children. *Journal of Nepal Paediatric Society* 2022; 42(2): 12–5.
35. Atwan ZW. GAPDH spike RNA as an alternative for housekeeping genes in relative gene expression assay

- using real-time PCR. Bulletin of the National Research Centre 2020; 44: 1–8.
36. Barber RD, Harmer DW, Coleman RA, Clark BJ. GAPDH as a housekeeping gene: analysis of GAPDH mRNA expression in a panel of 72 human tissues. Physiological genomics 2005; 21(3): 389–95.
37. Abdul Maksoud RS, Rashad NM, Elsayed WS, Ali MA, Kamal NM, Zidan HE. Circulating miR 181a and miR 223 expression with the potential value of biomarkers for the diagnosis of systemic lupus erythematosus and predicting lupus nephritis. The Journal of Gene Medicine 2021; 23(5): e3326.
38. Roffel MP, Boudewijn IM, van Nijnatten JL, Faiz A, Vermeulen CJ, van Oosterhout AJ, et al. Identification of asthma-associated microRNAs in bronchial biopsies. European Respiratory Journal 2022; 59(3).
39. Zhou Y, Zhang T, Yan Y, You B, You Y, Zhang W, et al. MicroRNA-223-3p regulates allergic inflammation by targeting INPP4A. Brazilian Journal of Otorhinolaryngology 2021; 87(5): 591–600.
40. Liu J, Su B, Tao P, Yang X, Zheng L, Lin Y, et al. Interplay of IL-33 and IL-35 Modulates Th2/Th17 Responses in Cigarette Smoke Exposure HDM-Induced Asthma. Inflammation 2024; 47(1): 173–90.
41. Guiot J, Struman I, Louis E, Louis R, Malaise M, Njock M-S. Exosomal miRNAs in lung diseases: from biologic function to therapeutic targets. Journal of Clinical Medicine 2019; 8(9): 1345.
42. Roffel MP, Maes T, Brandsma C-A, van den Berge M, Vanaudenaerde BM, Joos GF, et al. MiR-223 is increased in lungs of patients with COPD and modulates cigarette smoke-induced pulmonary inflammation. American Journal of Physiology-Lung Cellular and Molecular Physiology 2021; 321(6): L1091–L104.
43. Bandi DP, Sudhakar U, Parthasarathy H, Rajamani SR, Krishnaswamy B. Expression dynamics of microRNA-223/Ras-associated binding protein 12 axis in Stage III/Grade B periodontal disease: A case-control analysis. Journal of Indian Society of Periodontology 2024; 28(1): 99–105.
44. Huang K, Lin Z, Ge Y, Chen X, Pan Y, Lv Z, et al. Immunomodulation of MiRNA-223-based nanoplateform for targeted therapy in retinopathy of prematurity. Journal of Controlled Release 2022; 350: 789–802.
45. Mirra D, Cione E, Spaziano G, Esposito R, Sorgenti M, Granato E, et al. Circulating microRNAs expression profile in lung inflammation: A preliminary study. Journal of Clinical Medicine 2022; 11(18): 5446.
46. Ernst LM, Mithal LB, Mestan K, Wang V, Mangold KA, Freedman A, et al. Umbilical cord miRNAs to predict neonatal early onset sepsis. PLoS One 2021; 16(5): e0249548.
47. Lee MJ, Planck SR, Choi D, Harrington CA, Wilson DJ, Dailey RA, et al. Non-specific orbital inflammation: current understanding and unmet needs. Progress in Retinal and Eye Research 2021; 81: 100885.

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