

MULTIFACETED ROLES OF SUPEROXIDE DISMUTASES (SODS) IN CELLULAR HOMEOSTASIS AND CANCER PROGRESSION: REDOX REGULATION AND THERAPEUTIC IMPLICATIONS

VIŠESTRUKE ULOGE SUPEROKSID-DISMUTAZA (SOD) U ĆELIJSKOJ HOMEOSTAZI
I PROGRESIJI KARCINOMA: REDOKS REGULACIJA I TERAPIJSKE IMPLIKACIJE

Duygu Aydemir¹, Nuriye Nuray Ulusu^{2,3}

¹Istanbul University, Institute of Child Health, Department of Pediatric Basic Sciences,
Division of Medical Genetics, 34093, İstanbul, Türkiye

²Koç University, School of Medicine, Department of Medical Biochemistry, Sariyer, 34450, İstanbul, Türkiye

³Koç University Research Center for Translational Medicine (KUTTAM), Sariyer, 34450, İstanbul, Türkiye

Summary

Superoxide dismutases (SODs) are critical metalloenzymes that regulate cellular redox homeostasis by catalysing the dismutation of superoxide radicals into hydrogen peroxide and oxygen, thereby mitigating oxidative stress. Comprising three isoforms – SOD1 (Cu/Zn-SOD), SOD2 (Mn-SOD), and SOD3 (ecSOD) – these enzymes are localised in distinct cellular compartments, including the cytosol, mitochondria, and extracellular matrix, respectively. SODs play pivotal roles in cellular signalling, metabolism, and protection against reactive oxygen species (ROS)-mediated damage. Dysregulation of SOD expression and activity is implicated in various pathological conditions, particularly cancer, where they influence tumour initiation, progression, metastasis, and therapy resistance. Elevated SOD1 and SOD2 levels often promote oncogenic signalling and tumour survival, whereas SOD3 exhibits context-dependent roles, balancing tumour suppression and progression. Additionally, SOD mimetics, notably manganese-based compounds such as Mn-porphyrins and Mn-salens, have emerged as promising therapeutic agents that selectively modulate oxidative stress in cancer cells, thereby enhancing the efficacy of chemotherapy and radiotherapy while protecting normal tissues. This review explores the multifaceted roles of SODs in cellular homeostasis, their involvement in cancer pathogenesis, and the therapeutic potential of SOD mimetics in redox-based cancer strategies.

Keywords: superoxide dismutases, ROS, cellular homeostasis, cancer progression, redox regulation, antioxidant systems, tumour microenvironment, oxidative stress, SOD1, SOD2, SOD3

Address for correspondence:

N. Nuray Ulusu
Koc University, School of Medicine
Rumelifeneri Yolu 34450, Sariyer, İstanbul, Turkey
Phone: +90 (212) 338 11 60
Fax: +90 (212) 338 11 68
e-mail: nulusu@ku.edu.tr

Kratak sadržaj

Superoksid-dismutaze (SOD) predstavljaju ključne metalo-enzime u održavanju ćelijske redoks homeostaze, katalizujući dismutaciju superoksidnih radikalnih u vodonik-peroksid i kiseonik, čime se ublažava oksidativni stres. Tri izoforme ovih enzima – SOD1 (Cu/Zn-SOD), SOD2 (Mn-SOD) i SOD3 (ecSOD) – lokalizovane su u različitim ćelijskim odeljcima, uključujući citosol, mitohondrije i vanćelijski matriks. SOD enzimi imaju sуштинsku ulogu u ćelijskoj signalizaciji, metabolizmu i zaštiti od oštećenja izazvanih reaktivnim kiseoničnim vrstama (ROS). Poremećaji u ekspresiji i aktivnosti SOD enzima su povezani sa brojnim patološkim stanjima, naročito sa kancerom, gde doprinose inicijaciji tumora, njegovoj progresiji, metastaziranju i razvoju rezistencije na terapiju. Povišena ekspresija SOD1 i SOD2 najčešće podstiče onkogenu signalizaciju i održavanje tumorskih ćelija, dok SOD3 pokazuje kontekstualno zavisne efekte, balansirajući između tumorske supresije i progresije. Dodatno, SOD mimeticci, naročito jedinjenja na bazi mangana kao što su Mn-porfirini i Mn-saleni, izdvajaju se kao perspektivni terapeutski agensi, jer selektivno modulišu oksidativni stres u ćelijama raka. Na taj način povećavaju efikasnost hemoterapije i radioterapije, uz istovremenu zaštitu normalnih tkiva. Ovaj rad razmatra višestruke funkcije SOD enzima u ćelijskoj homeostazi, njihovu ulogu u patogenezi kancera i terapijski potencijal SOD mimetika u okviru redoks-zasnovanih strategija lečenja.

Ključne reči: superoksid-dismutaze, ROS, ćelijska homeostaza, progresija kancera, redoks regulacija, antioksidativni sistemi, tumorsko mikrookruženje, oksidativni stres, SOD1, SOD2, SOD3

Duygu Aydemir, Assistant Professor
Istanbul University, Institute of Child Health
Istanbul Tip Fakültesi Çapa – 34093 Fatih/Istanbul
e-mail: duygu.aydemir@istanbul.edu.tr

Introduction

Superoxide dismutase (SOD, EC 1.15.1.1) enzymes are a group of metalloenzymes found in all eukaryotic and prokaryotic organisms that live on oxygen (1, 2). SOD enzymes can detoxify free radicals and play a pivotal role in managing oxidative stress (3). These enzymes primarily protect aerobic organisms from the toxic effects of reactive oxygen species (ROS). The mitochondria, essential for adenosine triphosphate (ATP) production, are the primary source of superoxide anions (4). SOD enzymes can catalyse the conversion of superoxide anion ($O_2^{\cdot-}$) into less harmful compounds, such as oxygen and hydrogen peroxide (H_2O_2), through a cyclic oxidation-reduction mechanism (5). There are various ways to produce H_2O_2 , but SOD enzymes mainly control H_2O_2 concentration (6). For instance, plants, bacteria, fungi, and most other organisms except primates have urate oxidase enzymes that catalyse the production of H_2O_2 (7). The best-known property of SOD enzymes is the antioxidant defence against oxidative stress via the scavenging of ROS (8). SOD enzymes regulate essential processes in cellular signalling and primary cell metabolism, lipid metabolism, and inflammation involved in cell growth, proliferation, differentiation, cell survival, and apoptosis (9, 10), lipid peroxidation, oxidation of low-density lipoprotein in macrophages, lipid droplet formation, and adhesion of inflammation (11), and mitochondrial dysfunction (12). SOD enzymes protect the cells from radical attack; there-

fore, they are accepted as a therapeutic agent against ROS-mediated diseases (13).

Discussion

SOD enzymes are recognised as the primary enzyme in the detoxification/scavenging of $O_2^{\cdot-}$, which is generated during cellular metabolism, environmental stresses, and programmed cell death (14–16). SOD enzymes may trigger endogenous antioxidant enzymes for the neutralisation of ROS molecule H_2O_2 in a variety of pathological health conditions, for instance, xenobiotic metabolism, metabolic diseases, atherosclerosis, hypertension, angiogenesis, diabetes, cancer, pulmonary hypertension, nephropathy, edema, brain disease (Table 1) (12), cardiovascular diseases, cancer, respiratory diseases, skin diseases, renal, ocular diseases, neurological diseases, gastrointestinal diseases (17–22). It has been investigated that SOD enzyme activity is significantly decreased in patients with androgenetic alopecia (23) and those with Fanconi's anaemia (24). SOD enzyme is a potential marker in patients with viral hepatitis E-induced liver failure. SOD enzyme level is increasing in patients with viral hepatitis E due to elevated oxidative stress during the pathogenesis of the disease (25).

SOD enzymes are localised in the cytosol, mitochondria, chloroplasts (26), nucleus, lysosomes, peroxisomes, extracellular matrix, cell surface, and extra-

Table 1 Classes, representative compounds, mechanistic roles, and therapeutic implications of superoxide dismutase (SOD) mimetics.

Class of SOD Mimetics	Representative Compounds	Mechanistic Role	Therapeutic Implications
Mn Porphyrins	MnTnHex-2-PyP ⁵⁺ , MnBuOE-2-PyP ⁵⁺ , HSJ-0017	Catalyse dismutation of superoxide and peroxy nitrite; promote intracellular ROS accumulation in tumour cells; interfere with DNA repair; exert dual radiosensitising and radioprotective effects	Enhance efficacy of radiotherapy and chemotherapy (breast, lung, melanoma, glioblastoma); reduce normal tissue toxicity; potential combinatorial use with platinum-based drugs
Mn Salens	EUK-134, EUK-189	Mimic both SOD and catalase activity; attenuate oxidative damage; induce cell cycle arrest and inhibit migration	Suppress tumour proliferation, particularly in breast cancer; potential to overcome drug resistance via redox modulation
Mitochondria-Targeted Quinones	MitoQ10	Localise to mitochondria via lipophilic cation; modulate mitochondrial ROS and trigger apoptosis and autophagy	Induce mitochondrial destabilisation and tumour cell death; synergistic effects with natural compounds (e.g., curcumin) in redox-sensitive cancers
Nitroxides	Mito-TEMPO	Act as mitochondria-specific SOD mimetics; scavenge mitochondrial superoxide; inhibit NLRP3 inflammasome	Modulate inflammatory signalling and tumour microenvironment; protective role in ischemia–reperfusion injury and inflammation-associated malignancies
Mangafodipir Derivatives	Mangafodipir, Calmangafodipir	Initially developed as MRI contrast agents; exhibit MnSOD-mimetic and iron-chelating activity; Calmangafodipir is engineered to reduce Mn toxicity	Enhance chemotherapy efficacy (oxaliplatin, platinum derivatives); protect against haematological toxicity and chemotherapy-induced peripheral neuropathy

cellular fluids (Table 1) (12). They require metals for catalytic activity and can be classified according to the type of metal cofactors they use. For instance, copper-zinc superoxide dismutase, also known as SOD1, is commonly found in eukaryotes (27), while the iron-manganese-containing superoxide dismutase enzyme is found in bacteria and chloroplasts (28). Ni-containing SOD enzymes are found in prokaryotes such as *Streptomyces* sp. (29). On the other hand, three SOD enzymes are found in plants with different metal cofactors: Fe-SOD, Cu-SOD, and Mn-SOD (30). Alternatively, SOD isoenzymes are named SOD1, SOD2, and SOD3, and their cellular localisation is also essential. SOD1 or Cu, Zn-SOD enzyme is generally accepted as the central enzyme (31). Found in the cytosol and mitochondrial intermembrane space (IMS), nucleus, and endoplasmic reticulum (32), lysosomes, and peroxisomes (12). Mn-SOD is SOD2, found in the mitochondrial matrix and mitochondrial inner membrane (33), whereas the one found in the extracellular matrix is named SOD3 (33, 34).

SOD enzymes convert two O_2^- into H_2O_2 , which is further catalysed into H_2O and O_2 by catalase (CAT) and glutathione peroxidase (GPx) enzymes. GPx and GST enzymes are antioxidant enzymes that require GSH to function, and an

increased GSSG/GSH ratio is the cell's primary biomarker for oxidative stress status. Altered function in the antioxidant defence system causes enhanced accumulation of ROS associated with various disease pathogeneses (35). Oxidative stress is described by the increased accumulation of ROS and decreased antioxidant defence in the cell. Decreased SOD levels lead to the accumulation of ROS, primarily O_2^- , within the cell, triggering nucleophilic reactions. Since O_2^- deprotonates phenols, alcohols, and thiols and hydrolyses esters, it phosphorylates various types of proteins, including PKC, PKD (protein kinase D), PKB (Akt) (protein kinase B), and mitogen-activated (MAPK) kinases, p42/44, p38, ERK, and PI3K (36). Excessive ROS, including O_2^- , attacks DNA, lipids, and proteins, causing DNA damage and lipid peroxidation (18). Most types of cancer elevate their intrinsic ROS levels to promote oncogenic pathways, including genetic alterations, metabolic reprogramming, and metastasis. Thus, regulating oxidative stress metabolism in the tumour microenvironment and peripheral tissues is vital for controlling oncogenesis and developing hemotherapeutic approaches (37). Since the SOD family is the central regulator of oxidative stress metabolism and antioxidant response, recent studies have highlighted the significant role of the SOD family in cancer progression (Figure 1) (38).

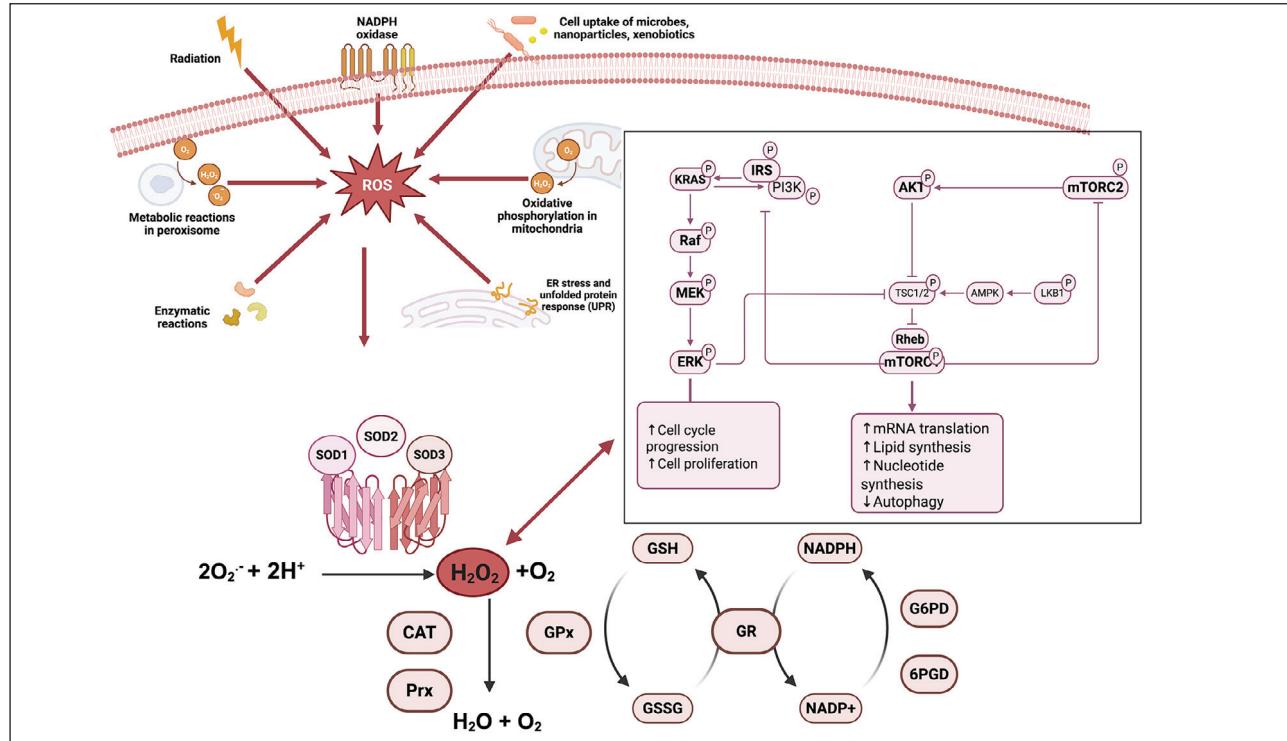


Figure 1 The role of Superoxide Dismutase (SOD) enzyme in the cell is associated with signalling pathways involved in cancer progression.

This figure illustrates the potential involvement of SOD enzymes in cancerous cells. SOD enzymes, which play a crucial role in metabolising reactive oxygen species (ROS), are present in both normal and cancer cells. In normal cells, SOD enzymes help maintain cellular redox balance by converting superoxide radicals (O_2^-) into less reactive molecules, such as hydrogen peroxide (H_2O_2) and oxygen. In contrast, in cancer cells, altered expression or activity of SOD enzymes may lead to an imbalance in ROS levels, contributing to genomic instability, tumorigenesis, and metastasis.

The role of the SOD enzymes in cancer pathogenesis

SOD1

SOD1 enzyme is a 32 kDa homodimeric metalloprotein containing Cu and Zn, and provides 80% of all SOD activity in the cell. SOD1 enzyme was discovered 50 years ago and is associated with the pathogenesis of various diseases, including cancer, neurological disorders, reduced fertility, ageing, metabolic disorders, and muscle wasting (39). SOD1 enzyme transcriptionally regulated, and this isoenzyme's activity is also controlled by posttranscriptional modifications (PTM). PTM can regulate conformation, function, and localisation (40). SOD1 has a controversial role in cancer progression because decreased SOD levels lead to an increase in ROS levels associated with oncogenesis. On the other hand, cancer cells require antioxidant enzymes, such as the SOD family, to prevent oxidative stress-induced cell damage and apoptosis (41). Increased SOD1 activity results in the H_2O_2 accumulation in the cell that triggers various signalling cascades, including AMP-activated protein kinase (AMPK), mitogen-activated protein kinase (MAPK), nuclear factor- κ B (NF- κ B), c-Jun N-terminal kinase (JNK), phosphatidylinositol 3-kinase-Akt (PI3K-Akt), Janus kinase-signal transducers and activator of transcription (JAK-STAT), activator protein-1 (AP-1), nuclear factor (erythroid-derived 2)-related factor 2 (Nrf2), hypoxia-inducible factor (HIF), Ras, and Rac involved in the progression, tumorigenesis, angiogenesis, and metastasis of cancer cells (42).

SOD1 plays a regulatory role in H_2O_2 -mediated signalling, and the overexpression of SOD1 activates the MAPK, JNK, and Akt pathways, as well as the AP-1, TNF- α , and JAK-STAT pathways involved in tumorigenesis (43). On the other hand, SOD1 localised in the nucleus regulates transcription of the stress response genes under enhanced oxidative stress in the cell. For instance, SOD1 induces glycolysis and suppresses oxidative phosphorylation (44). Overexpression of SOD1 has been reported in various types of cancers, such as non-small cell lung cancer (NSCLC), pancreatic cancer, liver cancer, brain cancer, nasopharyngeal carcinoma (NPC), leukaemia, and breast cancer (45). Knockdown or pharmaceutical inhibition of SOD1 blocks the proliferation of NSCLC, fibrosarcoma, pancreatic cancer, and breast cancer cells (46). SOD1 overexpression has been linked to cisplatin resistance in human ovarian cancer cells, and targeting SOD1 results in hemosensitivity in these cell lines (47).

SOD2

Human SOD2 (MnSOD) is a nuclear-encoded enzyme located in the mitochondria and controlled at transcriptional, translational, and post-translational levels. The SOD2 gene encodes an 88-kDa homotetrameric protein, and the enzyme's active site contains

manganese (48). Nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), specificity protein 1 (Sp1), activating protein-1 (AP-1), p53, and CCAAT binding protein (C/EBP) directly regulate SOD2 transcription under oxidative stress conditions via directly binding its promoter. On the other hand, pro-inflammatory cytokines (TNF- α , IL-1 β , IFN- γ), growth factors (TGF- β), fibroblast and epidermal growth factor, platelet-derived growth factor, endotoxins, extracellular factors (lipopolysaccharide, c-AMP, UV), nitric oxide, ionising radiation, viral infection, hypoxia, bacterial infection, and anticancer drugs (tamoxifen, paclitaxel, vinblastine, vincristine) can stimulate SOD2 transcription (38, 48–52).

Mitochondria are the primary source of oxidative stress in the cell due to oxidative phosphorylation; thus, SOD2 plays a vital role in antioxidant defence in both healthy and cancer cells. Homozygous knockout of SOD2 causes lethality in mice; however, homozygous knockout models of SOD1 and SOD3 have reportedly survived, according to the literature (53). SOD2 overexpression enhances mitochondrial function by scavenging O_2^{*-} produced with the mitochondria (54). The role of SOD2 in cancer pathogenesis is complex, as changes in SOD2 expression and activity are tumour-type dependent. Decreased SOD2 expression is frequently observed during tumour initiation; however, SOD2 levels increase during tumour progression and metastasis (55). Although increased SOD2 levels enable cells to cope with oxidative stress by scavenging O_2^{*-} , an SOD2-dependent increase in H_2O_2 leads to a shift in the cell's metabolism towards a H_2O_2 -dependent redox signalling status (56).

Increased SOD2 levels are associated with a 10-fold increase in the risk for pancreatic cancer; elevated H_2O_2 levels induced by SOD2 overexpression also led to increased GPx activity (57). Enhanced SOD2 induces tumorigenic and proangiogenic pathways such as VEGF, AKT, and HIF1 α in thyroid, pancreatic, colon, and breast cancers (58, 59). Since increased SOD2/ H_2O_2 , SOD2/CAT, and SOD2/GPx ratios are associated with tumour progression and metastasis in prostate, lung, and colon cancers, these ratios are considered valuable predictive biomarkers for the aforementioned cancer types (60). High SOD2 expression was linked to lymph node metastasis in oral squamous cell carcinoma (OSCC) (61).

SOD2 is downregulated in follicular thyroid cancer, and decreased expression of SOD2 is directly correlated with poor survival of patients having aggressive thyroid or adrenal cancer (62). The rs4880 polymorphism in exon 2 of SOD2 at position 16 is characterised by changing alanine (Ala) to valine (Val). This polymorphism leads to decreased mRNA expression and stability in SOD2, resulting in alterations in the enzyme's import into the mitochondria found in lung and oesophageal cancers (63). Reduced SOD2 transcription correlates with in-

creased mortality in hepatocellular carcinoma patients harbouring p53 mutation, indicating a tight relationship between p53 and SOD2 (64).

SOD3

SOD3 is the only antioxidant enzyme present in the extracellular matrix of restricted tissue and cell types; however, this enzyme has also been found in the nucleus and is trafficked via the endo-lysosomal system within the cell (65). The role of SOD3 enzymes in cancer development is multifaceted; they are involved in redox signalling, function as second messengers in regulatory pathways, contribute to maintaining genomic stability, and play a protective role in preventing carcinogenesis (66). The extracellular SOD (ecSOD; SOD3) has unique functions in cellular transduction, oxidative tumour microenvironment, tumour growth, metastasis, and recurrence (67). SOD3 is highly expressed in the lung, placenta, and cardiac endothelium, whereas it is moderately expressed in the pancreas, kidney, uterus, cartilage, brain, eye, skeletal muscle, and adipose tissue (68). After SOD3 is synthesised, it is bound to the cell surface proteoglycans via its positively charged heparin-binding domain (HBD). The HBD domain is cleaved by proteases, allowing SOD3 to be distributed into the extracellular milieu and the circulatory system (69). The primary function of the SOD3 is to control the radical levels in the cell via catalysing O_2^- into H_2O_2 , inhibiting the production of the $^{\bullet}OH$ via the Fenton and Haber Weiss reaction, and preventing the O_2^- -mediated oxidation of NO $^{\bullet}$ (65).

NO $^{\bullet}$ is produced by nitric oxide synthases (NOSs) and regulates the vessel relaxation in endothelial cells, neurotransmitter function, and macrophage and neutrophil functions. Firstly, the tissue-protective effects of SOD3 on cardiac tissue have been reported, and SOD3 administration has been shown to reduce cardiovascular damage, as documented in the literature (70). O_2^- -mediated peroxynitrite (ONOO $^-$) production from NO $^{\bullet}$ has adverse effects on cellular signalling; on the other hand, NO $^{\bullet}$ exerts both cancer-promoting and anticancer effects. ONOO $^-$ attacks lipids, DNA, and proteins via radical-mediated reactions that trigger cell death mechanisms. Enhanced ONOO $^-$ levels have been observed in stroke, cardiac arrest, myocardial infarction, diabetes, chronic shock, inflammatory diseases, neurodegenerative disorders, and cancer; thus, NO $^{\bullet}$ /ONOO $^-$ levels have a significant role in cancer pathogenesis (71).

Decreased SOD3 transcription and translation have been observed in patients with pancreatic cancer. This decrease in SOD3 is associated with a corresponding reduction in mean survival from 11.0 to 6.5 months in patients with pancreatic adenocarcinoma (72). Upregulation of SOD3 leads to the inhibition of

cell proliferation, clonogenic capacity, and invasion in a dose-dependent manner via decreasing VEGF and HIF-1 α protein levels. Since VEGF and HIF-1 α induce angiogenesis, SOD3 inhibits the blood flow into the tumour. On the other hand, SOD1 and SOD3 upregulation via Mirk/Dyrk1B kinase maintains cell growth by reducing ROS levels in the quiescent pancreatic cell lines Panc1 and SU86.86 in vitro (73). SOD3 upregulation decreased cell proliferation in melanoma cells via decreased IFNy and VEGF levels (74). SOD3-induced pathways in tumours connect vascular normalisation and T-cell diapedesis. Perivascular SOD3 prevents the oxidation of nitric oxide (NO), leading to increased endothelial cell (EC) NO levels, which inhibit prolyl hydroxylase domain (PHD) activity and cause the nuclear accumulation of HIF-2 α . HIF-2 α then upregulates vascular endothelial cadherin (VEC) and specific WNT ligands, which reduce vascular permeability and contribute to vascular normalisation (75). The upregulation of SOD3 reduces tumour growth and liver metastasis in colorectal cancer, indicating its potential as a diagnostic and prognostic marker in the treatment of colorectal cancer (76).

On the other hand, according to metabolic and proteomic studies, melanoma tumours recovered from chemotherapy showed enhanced SOD3 activity (77). SOD3 has protective effects on normal lung function, and downregulated SOD3 has been reported in lung cancer between stages I-IV. Overexpression of SOD3 reduced invasion and clonogenic survival by inhibiting NF- B in lung cancer (78).

SOD mimetics in diseases

Superoxide dismutase (SOD) mimetics are synthetic or low-molecular-weight compounds designed to replicate the activity of native SOD enzymes, which catalyse the dismutation of superoxide radicals into oxygen and hydrogen peroxide. These mimetics, particularly those based on manganese (Mn), have garnered significant interest due to their potential in mitigating oxidative stress-related pathologies, including cancer, neurodegenerative diseases, and inflammatory disorders. Mn-based SOD mimetics, including Mn-porphyrins, Mn-salen complexes, and Mn-pyridyl ligands, emphasise their structural versatility, catalytic mechanisms, and biological efficacy. Preclinical studies suggest these mimetics offer therapeutic advantages by modulating redox signalling and protecting tissues from oxidative damage, with some advancing toward clinical evaluation in various diseases, including head and neck, breast, brain, skin, and lymphoma cancers. On the other hand, Mn-based SOD mimetics enhance the efficacy of chemotherapy and radiotherapy, while also reducing tumour growth (79).

Table II Table II Isoenzyme-specific localisation, physiological roles, pathological involvement, and therapeutic implications of superoxide dismutases (SODs). SOD1, SOD2, and SOD3 are critical antioxidant enzymes that regulate cellular and extracellular redox balance by catalysing the dismutation of superoxide radicals into hydrogen peroxide and oxygen.

Isoenzyme	Cellular Localisation	Physiological Role	Role in Cancer and Pathology	Therapeutic Implications
SOD1 (Cu/Zn-SOD)	Cytosol, mitochondrial intermembrane space, nucleus, ER, peroxisomes, lysosomes	Detoxifies cytosolic superoxide radicals; regulates redox-sensitive signalling; balances glycolysis vs. oxidative phosphorylation	Overexpressed in lung, pancreatic, liver, breast cancers, leukaemia, and NPC; promotes tumour growth and survival via MAPK, NF- κ B, PI3K/Akt, JAK-STAT, HIF pathways; contributes to chemoresistance (cisplatin)	Targeting SOD1 sensitises tumours to chemotherapy; potential biomarker for therapy resistance; SOD1 inhibitors may block oncogenic signalling
SOD2 (Mn-SOD)	Mitochondrial matrix and inner membrane	Primary mitochondrial antioxidant defence; essential for survival; regulates oxidative phosphorylation	Tumour-type dependent: reduced in initiation, elevated in progression/metastasis; high SOD2/H2O2 promotes angiogenesis (VEGF, HIF-1 α , AKT); linked to prostate, thyroid, pancreatic, and colon cancer progression	Inhibition enhances sensitivity to chemotherapy (e.g., 5-FU in gastric cancer); SOD2 expression ratios (SOD2/GPx, SOD2/CAT) serve as prognostic biomarkers; SOD2 polymorphisms linked to cancer risk
SOD3 (Ec-SOD)	Extracellular matrix, circulation, nucleus (via endolysosomal trafficking)	Regulates extracellular ROS; preserves NO bioavailability; maintains vascular and tissue integrity	Downregulated in pancreatic and lung cancer (correlates with poor survival); overexpression suppresses invasion and angiogenesis; context-dependent tumour effects	Potential diagnostic/prognostic biomarker (lung, pancreatic, colorectal cancer); therapeutic upregulation may inhibit angiogenesis and metastasis; protective in normal tissues against oxidative injury

MnSOD mimetics in cancer therapy: a focus on compound classes and mechanistic insights

MnSOD mimetics represent a promising class of therapeutic agents in oncology due to their ability to selectively modulate redox homeostasis in tumour cells versus normal cells. Several subclasses of these mimetics, particularly Mn-based complexes and mitochondrial-targeted antioxidants, have demonstrated significant efficacy in preclinical cancer models. This section outlines the current understanding of the anti-cancer mechanisms and therapeutic potential of major MnSOD mimetics (Table II) (80).

Mn porphyrins

Among the MnSOD mimetics, manganese porphyrins are the most extensively studied, particularly for their potential to enhance the efficacy of radiotherapy and chemotherapy. MnTnHex-2-PyP⁵⁺ (MnTnHex) is a highly lipophilic Mn porphyrin that exhibits potent radio-sensitising effects in tumour cells. It improves the therapeutic efficacy of ionising radiation in breast cancer and melanoma models by promoting intracellular reactive oxygen species (ROS) accumulation and impairing DNA repair mechanisms. Additionally, MnTnHex has demonstrated synergism with cisplatin in non-small cell lung carcinoma (NSCLC) and renal car-

cinoma models, where it amplifies cytotoxicity and inhibits cancer cell migration, suggesting potential in combination chemotherapy regimens (79).

MnBuOE-2-PyP (MnBuOE) similarly exerts dual pro-oxidative and protective effects. In glioblastoma, ovarian, and lung cancer models, MnBuOE selectively increases oxidative stress in tumour cells, promoting apoptosis when used in conjunction with chemotherapeutic agents such as carboplatin, cisplatin, and tumour necrosis factor-related apoptosis-inducing ligand (TRAIL). MnBuOE has shown radio-protective effects in normal tissues, notably preserving hippocampal neurogenesis during cranial irradiation. This selective action enhances the therapeutic window of cancer treatment protocols (80).

HSJ-0017 is a recent Mn porphyrin compound combining anti-inflammatory and antitumor properties. In sarcoma models, HSJ-0017 augments the efficacy of chemotherapy and radiotherapy while concurrently mitigating treatment-induced toxicity. Its dual function underscores its value as a cytotoxic enhancer and a tissue-protective agent (81).

Mn salens

Mn salen complexes, such as EUK-134, mimic both SOD and catalase activity and have demonstrated

efficacy in cancer settings, particularly breast cancer. EUK-134 reduces intracellular superoxide and hydrogen peroxide levels, induces cell cycle arrest at the G2-M phase, and inhibits cell migration and adhesion. These actions collectively contribute to suppressed tumour proliferation and may counteract mechanisms underlying drug resistance. EUK-134's redox-modulating effects position it as a promising adjunct to existing therapies targeting redox-sensitive tumour pathways (81).

MitoQ10

Though not manganese-based, MitoQ10 functions as a mitochondria-targeted antioxidant with mechanistic parallels to MnSOD mimetics. It comprises a ubiquinone moiety linked to a triphenylphosphonium cation, facilitating mitochondrial accumulation. MitoQ10 induces mitochondrial destabilisation in cancer models, leading to apoptosis and autophagy. Its combination with agents such as curcumin further enhances tumoricidal activity, indicating potential utility in redox-sensitive cancer therapy (82).

Nitroxides

Nitroxide derivatives, particularly Mito-TEMPO, offer mitochondria-specific modulation of oxidative stress without containing manganese. Mito-TEMPO scavenges mitochondrial ROS and inhibits inflammatory signalling pathways such as the NOD-like receptor pyrin domain-containing 3 (NLRP3) inflammasome. These properties are particularly relevant in malignancies characterised by mitochondrial dysfunction and inflammation, positioning Mito-TEMPO as a potential therapeutic adjunct to modulate the tumour microenvironment and redox signalling (83).

Mangafodipir and Calmangafodipir

Initially developed as MRI contrast agents, Mangafodipir and its derivative Calmangafodipir have been repurposed for their MnSOD-mimetic properties. Calmangafodipir has been engineered to reduce manganese-associated toxicity through partial substitution with calcium. It has demonstrated the ability to enhance the efficacy of chemotherapy agents, such as oxaliplatin, while concurrently mitigating associated haematological toxicity and peripheral neuropathy. This dual action improves the therapeutic index of chemotherapeutic regimens and supports their development as chemoprotective adjuvants (84).

MnSOD mimetics, including Mn porphyrins, salens, mitochondria-targeted quinones, and related compounds, offer diverse and complementary mechanisms of action in cancer therapy. Their ability to selectively modulate oxidative stress within the tumour microenvironment presents a novel and multifaceted approach to enhancing therapeutic outcomes while minimising damage to normal tissues. Further clinical investigation is warranted to translate

these promising preclinical results into effective adjuncts for standard cancer treatments (85).

Conclusion

Superoxide dismutase (SOD) enzymes are central components of the cellular antioxidant defence system, critically involved in regulating redox balance by catalysing the dismutation of superoxide radicals into hydrogen peroxide. The three isoenzymes, SOD1, SOD2, and SOD3, each play distinct yet overlapping roles in maintaining cellular homeostasis and modulating oxidative stress in physiological and pathological contexts. Their dysregulation is closely associated with cancer initiation, progression, metastasis, and resistance to therapy. SOD1 and SOD2 are intimately linked to redox-sensitive oncogenic signalling pathways and contribute to tumour cell survival under oxidative stress. The SOD1-mTORC1 axis supports tumour adaptation to hypoxia and nutrient deprivation, while SOD2 expression correlates with poor prognosis and resistance to chemotherapeutic agents in several cancers. SOD3, although primarily protective in normal tissue, has context-dependent roles in tumour biology, influencing angiogenesis, invasion, and immune modulation.

Furthermore, the development of MnSOD mimetics, such as Mn porphyrins, salens, and mitochondria-targeted antioxidants, has opened new avenues in cancer therapy. These agents can selectively amplify oxidative stress in tumour cells while protecting normal tissues, enhancing the efficacy of chemotherapy and radiotherapy. Their dual action highlights the therapeutic potential of redox modulation in oncology. Overall, SOD enzymes not only serve as key modulators of oxidative stress but also emerge as critical biomarkers and therapeutic targets in cancer. Continued investigation into their regulatory mechanisms, interactions with cellular signalling networks, and pharmacological modulation through mimetics is essential for advancing redox-based strategies in precision cancer therapy.

Ethical approval

No ethical approval is required for this study.

Authors contributions

DA and NNU were responsible for conceptualisation, writing the original manuscript, and revising it.

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Conflict of interest statement

All the authors declare that they have no conflict of interest in this work.

References

- Wang Y, Branicky R, Noë A, Hekimi S. Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signaling. *J. Cell Biol* 2018; 217: 1915–28.
- Qiao K, Fang C, Chen B, Liu Z, Pan N, Peng H, Hao H, Xu M, Wu J, Liu S. Molecular characterization, purification, and antioxidant activity of recombinant superoxide dismutase from the Pacific abalone *Haliotis discus hanai* Ino. *World J Microbiol Biotechnol* 2020; 36:115.
- Borgstahl G, Oberley-Deegan R. Superoxide Dismutases (SODs) and SOD Mimetics. *Antioxidants* 2018; 7:156.
- Abdulrahman H, Dyary H, Mohammed R, Hamad D, Star F, Saeed N. Preventing free radical damage: The significance of including antioxidants in diet to strengthen immunity. *Open Vet J* 2024;14:1526.
- Bull C FJ. Steady-State Kinetic Studies of Superoxide Dismutases: Properties of the Iron Containing Protein from *Escherichia coli*. *J Am Chem Soc* 1985;107, 3295–304.
- Andrés CMC, Pérez de la Lastra JM, Juan CA, Plou FJ, Pérez-Lebeña E. Chemistry of Hydrogen Peroxide Formation and Elimination in Mammalian Cells, and Its Role in Various Pathologies. *Stresses* 2022; 2: 256–74.
- Wu XW, Lee CC, Muzny DM, Caskey CT. Urate oxidase: primary structure and evolutionary implications. *PNAS* 1989; 86: 9412–6.
- Banks CJ, Andersen JL. Mechanisms of SOD1 regulation by post-translational modifications. *Redox Biol* 2019; 26: 101270.
- Damiano S, Sozio C, La Rosa G, Guida B, Faraonio R, Santillo M, Mondola P. Metabolism Regulation and Redox State: Insight into the Role of Superoxide Dismutase 1. *Int J Mol Sci* 2020; 21: 6606.
- Palma FR, He C, Danes JM, Paviani V, Coelho DR, Gantner BN, Bonini MG. Mitochondrial Superoxide Dismutase: What the Established, the Intriguing, and the Novel Reveal About a Key Cellular Redox Switch. *Antioxid Redox Signal* 2020; 32: 701–14.
- Islam MN, Rauf A, Fahad FI, Emran T Bin, Mitra S, Olatunde A, Shariati MA, Rebezov M, Rengasamy KRR, Mubarak MS. Superoxide dismutase: an updated review on its health benefits and industrial applications. *Crit Rev Food Sci Nutr* 2022; 62: 7282–300.
- Fukai T, Ushio-Fukai M. Superoxide Dismutases: Role in Redox Signaling, Vascular Function, and Diseases. *Antioxid Redox Signal* 2011; 15: 1583–606.
- Younus H. Therapeutic potentials of superoxide dismutase. *Int J Health Sci (Qassim)* 2018; 12: 88–93.
- Reddy VN, Kasahara E, Hiraoka M, Lin L-R, Ho Y-S. Effects of variation in superoxide dismutases (SOD) on oxidative stress and apoptosis in lens epithelium. *Exp Eye Res* 2004; 79: 859–68.
- Nemmiche S. Oxidative Signaling Response to Cadmium Exposure. *Toxicological Sciences* 2016; 156(1): 4–10.
- Das K, Roychoudhury A. Reactive oxygen species (ROS) and response of antioxidants as ROS-scavengers during environmental stress in plants. *Front Environ Sci* 2014; 2.
- Ulusu NN, Sahilli M, Avci A, et al. Pentose phosphate pathway, glutathione-dependent enzymes and antioxidant defense during oxidative stress in diabetic rodent brain and peripheral organs: effects of stobadine and vitamin E. *Neurochem Res* 2003; 28(6): 815–23.
- Aydemir D, Ulusu NN. Comment on the: Molecular mechanism of CAT and SOD activity change under MPA-CdTe quantum dots induced oxidative stress in the mouse primary hepatocytes (*Spectrochim Acta A Mol Biomol Spectrosc* 2020; 229: 117792).
- Aydemir D, Malik AN, Kulac I, Basak AN, Lazoglu I, Ulusu NN. Impact of the Amyotrophic Lateral Sclerosis Disease on the Biomechanical Properties and Oxidative Stress Metabolism of the Lung Tissue Correlated With the Human Mutant SOD1G93A Protein Accumulation. *Front Bioeng Biotechnol* 2022; 10.
- Aydemir D, Ulusu NN. Importance of the serum biochemical parameters as potential biomarkers for rapid diagnosis and evaluating preclinical stage of ALS. *Med Hypotheses* 2020; 141: 109736.
- Aydemir D, Surucu S, Basak AN, Ulusu NN. Evaluation of the Hematological and Serum Biochemistry Parameters in the Pre-Symptomatic and Symptomatic Stages of ALS Disease to Support Early Diagnosis and Prognosis. *Cells* 2022; 11: 3569.
- Rosa AC, Corsi D, Cavi N, Bruni N, Dosio F. Superoxide Dismutase Administration: A Review of Proposed Human Uses. *Molecules* 2021; 26: 1844.
- Tulić L, Tulić I, Stojnić J, Bila J, Vuković Ž, Kotlica B. Different doses of recombinant FSH and determining parameters of oxidative stress. *J Med Biochem* 2024; 43 (1): 219–25.
- Yoshimitsu K, Kobayashi Y, Usui T. Decreased Superoxide Dismutase Activity of Erythrocytes and Leukocytes in Fanconi's Anemia. *Acta Haematol* 1984; 72: 208–10.
- He Y, Wang F, Yao N, Wu Y, Zhao Y, Tian Z. Serum superoxide dismutase level is a potential biomarker of disease prognosis in patients with HEV-induced liver failure. *BMC Gastroenterol* 2022; 22: 14.
- Grace SC. Phylogenetic distribution of superoxide dismutase supports an endosymbiotic origin for chloroplasts and mitochondria. *Life Sci* 1990; 47: 1875–86.
- Fetherolf MM, Boyd SD, Taylor AB, Kim HJ, Wohlschlegel JA, Blackburn NJ, Hart PJ, Winge DR, Winkler DD. Copper-zinc superoxide dismutase is activated through a sulfenic acid intermediate at a copper ion entry site. *Journal of Biological Chemistry* 2017; 292: 12025–40.
- Ganini D, Petrovich RM, Edwards LL, Mason RP. Iron incorporation into MnSOD A (bacterial Mn-dependent superoxide dismutase) leads to the formation of a peroxidase/catalase implicated in oxidative damage to

bacteria. *Biochimica et Biophysica Acta (BBA) - General Subjects* 2015; 1850: 1795–805.

29. Youn HD, Kim EJ, Roe JH, Hah YC, Kang S-O. A novel nickel-containing superoxide dismutase from *Streptomyces* spp. *Biochemical Journal* 1996; 318: 889–96.

30. Gallie DR, Chen Z. Chloroplast-localized iron superoxide dismutases FSD2 and FSD3 are functionally distinct in *Arabidopsis*. *PLoS One* 2019; 14:e0220078.

31. ciskalska M, Ołdakowska M, Marek G, Milnerowicz H. Changes in the Activity and Concentration of Superoxide Dismutase Isoenzymes (Cu/Zn SOD, MnSOD) in the Blood of Healthy Subjects and Patients with Acute Pancreatitis. *Antioxidants* 2020; 9: 948.

32. Kawamata H, Manfredi G. Import, Maturation, and Function of SOD1 and Its Copper Chaperone CCS in the Mitochondrial Intermembrane Space. *Antioxid Redox Signal* 2010; 13: 1375–84.

33. Zelko IN, Mariani TJ, Folz RJ. Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. *Free Radic Biol Med* 2002; 33: 337–49.

34. Wert KJ, Velez G, Cross MR, Wagner BA, Teoh-Fitzgerald ML, Buettner GR, McAnany JJ, Olivier A, Tsang SH, Harper MM, et al. Extracellular superoxide dismutase (SOD3) regulates oxidative stress at the vitreoretinal interface. *Free Radic Biol Med* 2018; 124: 408–19.

35. Kuninaka S, Ichinose Y, Koja K, Toh Y. Suppression of manganese superoxide dismutase augments sensitivity to radiation, hyperthermia and doxorubicin in colon cancer cell lines by inducing apoptosis. *Br J Cancer* 2000; 83: 928–34.

36. Jin Q, Jhun BS, Lee SH, Lee J, Pi Y, Cho YH, Baik HH, Kang I. Differential regulation of phosphatidylinositol 3-kinase/Akt, mitogen-activated protein kinase, and AMP-activated protein kinase pathways during menadione-induced oxidative stress in the kidney of young and old rats. *Biochem Biophys Res Commun* 2004; 315: 555–61.

37. Nogueira V, Hay N. Molecular Pathways: Reactive Oxygen Species Homeostasis in Cancer Cells and Implications for Cancer Therapy. *Clinical Cancer Research* 2013; 19: 4309–14.

38. Hileman EA, Achanta G, Huang P. Superoxide dismutase: an emerging target for cancer therapeutics. *Expert Opin Ther Targets* 2001; 5: 697–710.

39. Zhang Y, Unnikrishnan A, Deepa SS, Liu Y, Li Y, Ikeno Y, Sosnowska D, Van Remmen H, Richardson A. A new role for oxidative stress in aging: The accelerated aging phenotype in Sod1^{−/−} mice is correlated to increased cellular senescence. *Redox Biol* 2017; 11: 30–7.

40. Eleutherio ECA, Silva Magalhães RS, de Araújo Brasil A, Monteiro Neto JR, de Holanda Paranhos L. SOD1, more than just an antioxidant. *Arch Biochem Biophys* 2021; 697: 108701.

41. Che M, Wang R, Li X, Wang H-Y, Zheng XFS. Expanding roles of superoxide dismutases in cell regulation and cancer. *Drug Discov Today* 2016; 21: 143–9.

42. Li X, Chen Y, Zhao J, Shi J, Wang M, Qiu S, Hu Y, Xu Y, Cui Y, Liu C, et al. The Specific Inhibition of SOD1 Selectively Promotes Apoptosis of Cancer Cells via Regulation of the ROS Signaling Network. *Oxid Med Cell Longev* 2019; 2019: 1–21.

43. Oakley FD, Abbott D, Li Q, Engelhardt JF. Signaling Components of Redox Active Endosomes: The Redoxosomes. *Antioxid Redox Signal* 2009; 11: 1313–33.

44. Reddi AR, Culotta VC. SOD1 Integrates Signals from Oxygen and Glucose to Repress Respiration. *Cell* 2013; 152: 224–35.

45. Liu S, Li B, Xu J, Hu S, Zhan N, Wang H, Gao C, Li J, Xu X. SOD1 Promotes Cell Proliferation and Metastasis in Non-small Cell Lung Cancer via an miR-409-3p/SOD1/SETDB1 Epigenetic Regulatory Feedforward Loop. *Front Cell Dev Biol* 2020; 8.

46. Tsang CK, Chen M, Cheng X, Qi Y, Chen Y, Das I, Li X, Vallat B, Fu L-W, Qian C-N, et al. SOD1 Phosphorylation by mTORC1 Couples Nutrient Sensing and Redox Regulation. *Mol Cell* 2018; 70: 502–515.e8.

47. Kim JW, SHYJWM. Knock-down of superoxide dismutase 1 sensitizes cisplatin-resistant human ovarian cancer cells. *Anticancer Res* 2010; 30: 2577–81.

48. Ludwig ML, Metzger AL, Patridge KA, Stallings WC. Manganese superoxide dismutase from *Thermus thermophilus*. *J Mol Biol* 1991; 219: 335–58.

49. Harris CA, DKH-MBKMCKSDELB. Manganese superoxide dismutase is induced by IFN-gamma in multiple cell types. Synergistic induction by IFN-gamma and tumor necrosis factor or IL-1. *J Immunol* 1991; 147: 149–54.

50. Maehara K, Oh Hashi K, Isobe K. Early growth responsive 1 dependent manganese superoxide dismutase gene transcription mediated by platelet derived growth factor. *The FASEB Journal* 2001; 15: 2025–6.

51. Warner BB, Burhans MS, Clark JC, Wispe JR. Tumor necrosis factor-alpha increases Mn-SOD expression: protection against oxidant injury. *Am J Physiol Lung Cell Mol Physiol* 1991; 260:296–301.

52. Strålin P, Marklund SL. Multiple cytokines regulate the expression of extracellular superoxide dismutase in human vascular smooth muscle cells. *Atherosclerosis* 2000; 151: 433–41.

53. Li Y, Huang T-T, Carlson EJ, Melov S, Ursell PC, Olson JL, Noble LJ, Yoshimura MP, Berger C, Chan PH, et al. Dilated cardiomyopathy and neonatal lethality in mutant mice lacking manganese superoxide dismutase. *Nat Genet* 1995; 11: 376–81.

54. Lee S, Van Remmen H, Csete M. Sod2 overexpression preserves myoblast mitochondrial mass and function, but not muscle mass with aging. *Aging Cell* 2009; 8: 296–310.

55. Kim Y, Gupta Vallur P, Phaëton R, Mythreye K, Hempel N. Insights into the Dichotomous Regulation of SOD2 in Cancer. *Antioxidants* 2017; 6: 86.

56. Hempel N, Melendez JA. Intracellular redox status controls membrane localization of pro- and anti-migratory signaling molecules. *Redox Biol* 2014; 2: 245–50.

57. Ansenberger-Fricano K, Ganini D, Mao M, Chatterjee S, Dallas S, Mason RP, Stadler K, Santos JH, Bonini MG. The peroxidase activity of mitochondrial superoxide dismutase. *Free Radic Biol Med* 2013; 54: 116–24.

58. Pringle DR, Yin Z, Lee AA, Manchanda PK, Yu L, Parlow AF, Jarjoura D, La Perle KMD, Kirschner LS. Thyroid-specific ablation of the Carney complex gene, PRKAR1A, results in hyperthyroidism and follicular thyroid cancer. *Endocr Relat Cancer* 2012; 19: 435–46.

59. Cen J, Zhang L, Liu F, Zhang F, Ji B-S. Long Term Alteration of Reactive Oxygen Species Led to Multidrug Resistance in MCF 7 Cells. *Oxid Med Cell Longev* 2016; 7053451, 15.

60. Miar A, Hevia D, Muñoz-Cimadevilla H, Astudillo A, Velasco J, Sainz RM, Mayo JC. Manganese superoxide dismutase (SOD2/MnSOD)/catalase and SOD2/GPx1 ratios as biomarkers for tumor progression and metastasis in prostate, colon, and lung cancer. *Free Radic Biol Med* 2015; 85: 45–55.

61. Yu B, Cao W, Zhang C, Xia R, Liu J, Yan M, Chen W. Prediction of lymph node metastasis in oral squamous cell carcinoma based on protein profile. *Expert Rev Proteomics* 2019; 16: 363–373.

62. Ashtekar A, Huk D, Magner A, La Perle KMD, Boucail L, Kirschner LS. Alterations in Sod2-Induced Oxidative Stress Affect Endocrine Cancer Progression. *J Clin Endocrinol Metab* 2018; 103: 4135–45.

63. Kang SW. Superoxide dismutase 2 gene and cancer risk: evidence from an updated meta-analysis. *Int J Clin Exp Med* 2015; 8: 14647–55.

64. Wang R, Yin C, Li X-X, Yang X-Z, Yang Y, Zhang M-Y, Wang H-Y, Zheng XFS. Reduced SOD2 expression is associated with mortality of hepatocellular carcinoma patients in a mutant p53-dependent manner. *Aging* 2016; 8: 1184–200.

65. Griess B, Tom E, Domann F, Teoh-Fitzgerald M. Extracellular superoxide dismutase and its role in cancer. *Free Radic Biol Med* 2017; 112: 464–479.

66. Zheng M, Liu Y, Zhang G, Yang Z, Xu W, Chen Q. The Applications and Mechanisms of Superoxide Dismutase in Medicine, Food, and Cosmetics. *Antioxidants* (2023) 12:1675. doi: 10.3390/antiox12091675

67. Griess B, Tom E, Domann F, Teoh-Fitzgerald M. Extracellular superoxide dismutase and its role in cancer. *Free Radic Biol Med* 2017; 112: 464–79.

68. Marklund SL. Extracellular superoxide dismutase in human tissues and human cell lines. *Journal of Clinical Investigation* 1984; 74: 1398–403.

69. Olsen DAa, Petersen S V, Oury TD, Valnickova Z, Thøgersen IB, Kristensen T, Bowler RP, Crapo JD, Enghild JJ. The Intracellular Proteolytic Processing of Extracellular Superoxide Dismutase (EC-SOD) is a Two-step Event. *Journal of Biological Chemistry* 2004; 279: 22152–7.

70. Laukkonen MO. Extracellular Superoxide Dismutase: Growth Promoter or Tumor Suppressor? *Oxid Med Cell Longev* 2016; 2016, 3612589, 9.

71. Vahora H, Khan MA, Alalami U, Hussain A. The Potential Role of Nitric Oxide in Halting Cancer Progression Through Chemoprevention. *J Cancer Prev* 2016; 21: 1–12.

72. O'Leary BR, Fath MA, Bellizzi AM, Hrabe JE, Button AM, Allen BG, Case AJ, Altekruse S, Wagner BA, Buettner GR, et al. Loss of SOD3 (EcSOD) Expression Promotes an Aggressive Phenotype in Human Pancreatic Ductal Adenocarcinoma. *Clinical Cancer Research* 2015; 21: 1741–51.

73. Deng X, Ewton DZ, Friedman E. Mirk/Dyrk1B Maintains the Viability of Quiescent Pancreatic Cancer Cells by Reducing Levels of Reactive Oxygen Species. *Cancer Res* 2009; 69: 3317–24.

74. Kim S-H, Kim M-O, Gao P, Youm C-A, Park H, Lee S-R, Kim K-S, Suh J-G, Lee H-T, Park B-J, et al. Overexpression of Extracellular Superoxide Dismutase (EC-SOD) in Mouse Skin Plays a Protective Role in DMBA/TPA-Induced Tumor Formation. *Oncol Res* 2005; 15(7-8): 333–41.

75. Martínez-Rey D, Carmona-Rodríguez L, Fernández-Aceñero MJ, Mira E, Mañes S. Extracellular Superoxide Dismutase, the Endothelial Basement Membrane, and the WNT Pathway: New Players in Vascular Normalization and Tumor Infiltration by T-Cells. *Front Immunol* 2020; 30:11: 579552.

76. Wang D, Chen M, Tao Z, Du J, Tian K, Chen Z, Yu B, Chen Y, Lv L. Overexpression of Extracellular Superoxide Dismutase 3 Inhibits Cancer Cell Growth and Migration in Colorectal Cancer. *Turk J Gastroenterol* 2024; 35(6): 465–74.

77. Morvan D, Demidem A. Metabolomics and transcriptomics demonstrate severe oxidative stress in both localized chemotherapy-treated and bystander tumors. *Biochimica et Biophysica Acta (BBA) - General Subjects* 2014; 1840: 1092–104.

78. Teoh-Fitzgerald MLT, Fitzgerald MP, Jensen TJ, Futscher BW, Domann FE. Genetic and Epigenetic Inactivation of Extracellular Superoxide Dismutase Promotes an Invasive Phenotype in Human Lung Cancer by Disrupting ECM Homeostasis. *Molecular Cancer Research* 2012; 10: 40–51.

79. Spasojević I, Chen Y, Noel TJ, Yu Y, Cole MP, Zhang L, Zhao Y, St. Clair DK, Batinic-Haberle I. Mn porphyrin-based superoxide dismutase (SOD) mimic, MnIIIITE-2-PyP5+, targets mouse heart mitochondria. *Free Radic Biol Med* 2007; 42: 1193–200.

80. Yulyana Y, Tovmasyan A, Ho IA, Sia KC, Newman JP, Ng WH, Guo CM, Hui KM, Batinic-Haberle I, Lam PY. Redox-Active Mn Porphyrin-based Potent SOD Mimic, MnTnBuOE-2-PyP5+, Enhances Carbenoxolone-Mediated TRAIL-Induced Apoptosis in Glioblastoma Multiforme. *Stem Cell Rev Rep* 2016; 12:140–55.

81. Lawler JM, Kunst M, Hord JM, Lee Y, Joshi K, Botchlett RE, Ramirez A, Martinez DA. EUK-134 ameliorates nNOS μ translocation and skeletal muscle fiber atrophy during short-term mechanical unloading. *Am J Physiol Regul Integr Comp Physiol* 2014; 1; 306(7): 470–82.

82. McLachlan J, Beattie E, Murphy MP, Koh-Tan CHH, Olson E, Beattie W, Dominiczak AF, Nicklin SA, Graham

D. Combined therapeutic benefit of mitochondria-targeted antioxidant, MitoQ10, and angiotensin receptor blocker, losartan, on cardiovascular function. *J Hypertens* 2014; 32: 555–64.

83. Patel K, Chen Y, Dennehy K, Blau J, Connors S, Mendonca M, Tarpey M, Krishna M, Mitchell JB, Welch WJ, et al. Acute antihypertensive action of nitroxides in the spontaneously hypertensive rat. *J Physiol Regul Integr Comp Physiol* 2006; 290(1): 37–43.

84. Karlsson JOG, Ignarro LJ, Lundström I, Jynge P, Almén T. Calmangafodipir [Ca4Mn(DPDP)5], mangafodipir (MnDPDP) and MnPLED with special reference to their SOD mimetic and therapeutic properties. *Drug Discov Today* 2015; 20: 411–21.

85. Grujicic J, Allen AR. MnSOD Mimetics in Therapy: Exploring Their Role in Combating Oxidative Stress-Related Diseases. *Antioxidants* 2024; 13: 1444.

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