

## THE IMPORTANCE OF POLYMORPHISMS OF REGULATORY AND CATALYTIC ANTIOXIDANT PROTEINS IN CHRONIC KIDNEY DISEASE

## ZNAČAJ POLIMORFIZAMA REGULATORNIH I KATALITIČKIH ANTIOKSIDATIVNIH PROTEINA U HRONIČNOJ BUBREŽNOJ SLABOSTI

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### Abstract

Both excessive production of reactive oxygen species (ROS) and impaired antioxidant function are found in patients with chronic kidney disease (CKD). Therefore, individual susceptibility towards CKD can be induced by functional variations of genes encoding antioxidant regulatory (nuclear factor erythroid 2 – related factor 2 (Nrf2)) and catalytic (superoxide dismutase (SOD2) and glutathione peroxidase (GPX1)) proteins. Several types of single nucleotide polymorphisms (SNPs) have been found within the genes encoding these proteins, with Nrf2 (-617C/A), SOD2 (*Ala16Val*) and GPX1 (*Pro198Leu*) conferring impaired catalytic activity. The most unexplored gene polymorphism in CKD susceptibility, progression and survival, with only two original studies published, is the Nrf2 (-617C/A) polymorphism. The results of these studies showed that there was no individual impact of this polymorphism on the susceptibility towards end stage renal disease (ESRD) development, oxidative phenotype and mortality. However, Nrf2 had a significant role in ESRD risk and survival, when combined with other antioxidant genes. The results regarding the impact of SOD2 (*Ala16Val*) and GPX1 (*Pro198Leu*) polymorphisms on either CKD or ESRD are still inconclusive. Namely, some studies showed that patients having variant SOD2 (*Val*) or GPX1 (*Leu*) allele were at increased risk of CKD development and progression, while other studies reported only weak or no association between these polymorphisms and CKD. Surprisingly, the only study that reported an association of GPX1 polymorphism with overall/cardiovascular survival in ESRD patients showed a significant impact of low activity GPX1 (*Leu/Leu*) genotype on better survival. In this review, we comprehensively and critically appraise the literature on these polymorphisms related to oxidative stress in CKD patients, in order to identify gaps and provide recommendations for further clinical research and translation. New developments in the field of antioxidant polymorphisms in CKD patients could lead to better stratification of CKD patients, based on a prognostic antioxidant gene panel, and provide a more personalised medicine approach for the need of antioxidant therapy in these patients.

### Keywords:

CKD,  
Nrf2,  
SOD2,  
GPX1,  
gene polymorphisms

## Sažetak

Hroničnu bubrežnu slabost (HBS) karakterišu prekomerno stvaranje slobodnih radikala i smanjenje antioksidativne zaštite. Stoga razlike u individualnoj podložnosti HBS mogu da leže u funkcionalnim varijacijama gena koji kodiraju regulatorne i katalitičke antioksidativne proteine, poput *Nrf2*, superoksid dizmutaze (*SOD*) i glutation-peroksidaze (*GPX*). Identifikovano je više različitih polimorfizama u okviru gena koji kodiraju ove enzime, poput *Nrf2* (-617C/A), *SOD2* (*Ala16Val*) and *GPX1* (*Pro198Leu*) i koji dovode do promena u njihovoj aktivnosti. Do sada najmanje ispitan polimorfizam od navedenih u podložnosti, progresiji bolesti i preživljavanju u HBS je *Nrf2* (-617C/A). Rezultati dve studije koje su ispitivale navedeni polimorfizam ukazuju da ne postoji individualna povezanost ovog polimorfizma sa rizikom za nastanak terminalne bubrežne slabosti (TBS), oksidativnim fenotipom ni preživljavanjem. S druge strane, *Nrf2* je imao značajan uticaj na rizik za nastanak TBS i preživljavanje u kombinaciji sa polimorfizmima drugih antioksidativnih gena. Rezultati istraživanja koji se odnose na povezanost *SOD2* (*Ala16Val*) i *GPX1* (*Pro198Leu*) polimorfizama i HBS su kontradiktorni. S jedne strane, pokazano je da pacijenti koji su nosioci manje aktivnih *SOD2* (*Val*) ili *GPX1* (*Leu*) alela imaju veći rizik za razvoj i progresiju HBS. Ipak, druge studije ukazuju na odustvo povezanosti ovih polimorfizama sa HBS. Rezultati jedinog istraživanja, u kome je ispitivana povezanost *GPX1* polimorfizma i preživljavanja (ukupnog i kardiovaskularnog) kod pacijenata sa TBS, pokazali su značajan uticaj *GPX1* (*Leu/Leu*) genotipa na duže preživljavanje. Rezultati prikazani u ovom radu mogu biti od značaja kako bi se dale preporuke za dalja klinička i translaciona istraživanja. Razvoj istraživanja u oblasti polimorfizama antioksidativnih gena kod pacijenata sa HBS-om može doprineti boljoj stratifikaciji ovih pacijenata na osnovu panela antioksidativnih gena. Takav panel bi mogao da bude dalje korišćen u kontekstu predviđanja preživljavanja CKD pacijenata, što bi takođe doprinelo razvijanju personalizovanog pristupa u njihovom lečenju, uključujući primenu ciljane antioksidativne terapije.

### Ključne reči:

HBS,  
*Nrf2*,  
*SOD2*,  
*GPX1*,  
geniski polimorfizmi

## Introduction

Chronic kidney disease (CKD) is a multifactorial disorder which is comprised of a broad range of adverse outcomes, accompanied with poor quality of life and high medical costs. CKD is a syndrome characterised by structural and/or functional kidney abnormalities, which persist for more than 3 months, or decreased glomerular filtration rate (GFR) below 60 mL/min/1.73 m<sup>2</sup>, irrespective of the underlying cause (1). With regard to GFR, CKD is classified into five stages (G1 - G5). The final stage of CKD is called end stage renal disease (ESRD), which occurs when GFR drops below 15 mL/min per/1.73 m<sup>2</sup> (stage G5) (1). At this point, kidney function is severely compromised and kidney replacement therapy is required. Therefore, CKD and its final stage, ESRD, are accompanied with excessive morbidity and mortality, mostly due to the cardiovascular complications and cancer (2-4). Oxidative stress has been demonstrated as one of the main contributing factors to CKD pathogenesis, as well as its long term complications (5-9).

Oxidative stress can be described as a disturbance in the balance between the prooxidants and antioxidants, where prooxidant insults predominate (10, 11). Previously, oxidative stress was conceptualized as pertaining to molecular damage (10). However, new findings paved the way towards the understanding of oxidative stress as relevant for redox signalling, as well (11). Both excessive production of reactive oxygen species (ROS) and impaired antioxidant function are frequently reported in CKD patients.

Several factors potentiate oxidative burden in CKD. Some of these factors include the following: accumulated uremic toxins, iron therapy, mitochondrial dysfunction, endoplasmic reticulum stress, as well as the increase in oxidative enzymes activity (myeloperoxidase, nicotinamide adenine dinucleotide phosphate-oxidases, xanthine oxidase) and endothelial nitric oxide synthase (eNOS) uncoupling (8, 12-17). Moreover, in ESRD patients, the increased generation of ROS is strongly mediated by haemodialysis treatment itself, mostly due to membrane bio-incompatibility and endotoxin (LPS) release (17, 18). On the other hand, antioxidant systems, including non-enzymatic (thiol, vitamin C, vitamin E), as well as enzymatic systems (superoxide dismutase (SOD), glutathione peroxidase (GPX), catalase (CAT)) are both impaired or deficient in patients with CKD (19-22). The antioxidant loss in CKD patients may be a consequence of the therapy with diuretics, illness-specific diet, as well as decreased intestinal absorption (23). Additionally, the haemodialysis in ESRD patients leads to solute loss, including loss of hydrophilic non-enzymatic antioxidants (19, 24, 25). The excessive production of free radicals induces oxidative (OH<sup>-</sup>), nitrosative (ONOO<sup>-</sup>) and chlorinative (OCl<sup>-</sup>) modifications of proteins, lipids and DNA, resulting in cellular and tissue injury (17, 22, 26-29). Indeed, heightened levels of oxidation markers of proteins, lipids and DNA damage have all been consistently reported in studies comparing clinical CKD population with healthy controls (30, 31). Oxidative stress is a mediator of progressive deterioration of kidney function, as well as of cardiovascular, neurological and

other complications in these patients: it is involved in the pathophysiology of endothelial dysfunction and atherosclerosis, hypertension, reduced erythrocyte half-life, and inflammation (7, 9, 32).

Excessive oxidative stress in patients with CKD could be related to specific genetic patterns (17). Namely, genetic polymorphisms of regulatory (nuclear factor erythroid 2-related factor 2 (Nrf2)) and catalytic antioxidant proteins (SOD2 and GPX1) result in alteration in their proteins activity profile, hence affecting individuals antioxidant capacity (33–36). In this comprehensive review, the focus was on the studies examining significance and roles of single nucleotide polymorphisms (SNPs) of Nrf2, SOD2 and GPX1 genes in CKD susceptibility and prognosis, outlining their importance and potential for clinical application.

## NRF2 Polymorphism and CKD

Nrf2 is a cytoprotective transcription factor which regulates basal activity, as well as the coordination of induction of a broad range of genes encoding antioxidant and detoxifying enzymes (SOD, CAT, GPX, glutathione S-transferase (GST), thyroredoxin, etc.) (30, 37–41). It binds to a Kelch-like ECH-associated protein 1 (Keap1) in the cytoplasm (39, 42, 43). Keap1, in turn, promotes the ubiquitination and proteasomal degradation of Nrf2 (38, 42, 44, 45). Several reactive cysteine residues present in Keap1 can be modified by electrophilic and oxidative insults which leads to the dissociation of Nrf2 from Keap1 and its transport to the nucleus, where it binds to the regulatory sequences, called antioxidant response elements (AREs) (17, 38, 41, 42, 44–46). AREs are located in the promoter region of an array of antioxidant and detoxifying genes (38, 41, 47). Therefore, Nrf2 has a critical position in cellular protection, orchestrating the expression of aforementioned genes (46, 48).

Increasing evidence from animal models supports

the crucial impact that Nrf2 has on renoprotection and kidney disease pathogenesis (30). Namely, animal models with 5/6 nephrectomy-induced CKD had a marked decline in nuclear Nrf2, in contrary to the expected Nrf2 activation and upregulation (46). This points to the impaired remnant kidney capacity to deal with the oxidative stress and inflammation (17, 46). Experimental data showing the influence of Nrf2 in diabetic nephropathy was also reported by other researchers (30, 49, 50). In a model of streptozotocin-induced diabetes, Jiang et al. found that Nrf2-knockout mice had higher ROS levels and consequently higher DNA oxidative damage in glomeruli that wild-type mice (17, 50). Furthermore, Nrf2 - knockout experimental animals developed a lupus-like autoimmune nephritis (46, 51, 52). In addition, in a model of ischemia - reperfusion injury, Nrf2 - knockout mice had a greater decline in renal function and shorter survival than wild - type mice (30, 53). On the other hand, the administration of Nrf2 activators, including resveratrol, curcumin, sulforaphane and bardoxolone methyl, can be protective against kidney dysfunction in CKD (30, 54). Therefore, studies using animal models with targeted deletion of Nrf2 have provided insights into the role of Nrf2 in kidney disease pathogenesis (30, 55). Subsequent studies investigating the genetic and molecular function of human Nrf2, including the relationship of Nrf2 polymorphisms with the CKD and its final stage - ESRD, have emerged recently (54 - 56).

Human Nrf2 is positioned on the cytogenetic band 2q31.2 of chromosome 2 (55, 57). Numerous SNPs have been found in the Nrf2 gene (33, 42, 55, 58). Of special importance is the functional Nrf2 rs6721961 polymorphism, located in the promoter region of the gene, which is characterised by a C > A substitution (33). It affects ARE-like promoter binding sites, attenuating the efficient binding of this transcription factor to AREs (33, 58). This results in a reduction in the Nrf2 transcription activity, and a consequent decrease in Nrf2-dependent antioxidant and detoxifying gene transcription (33). So far, only two

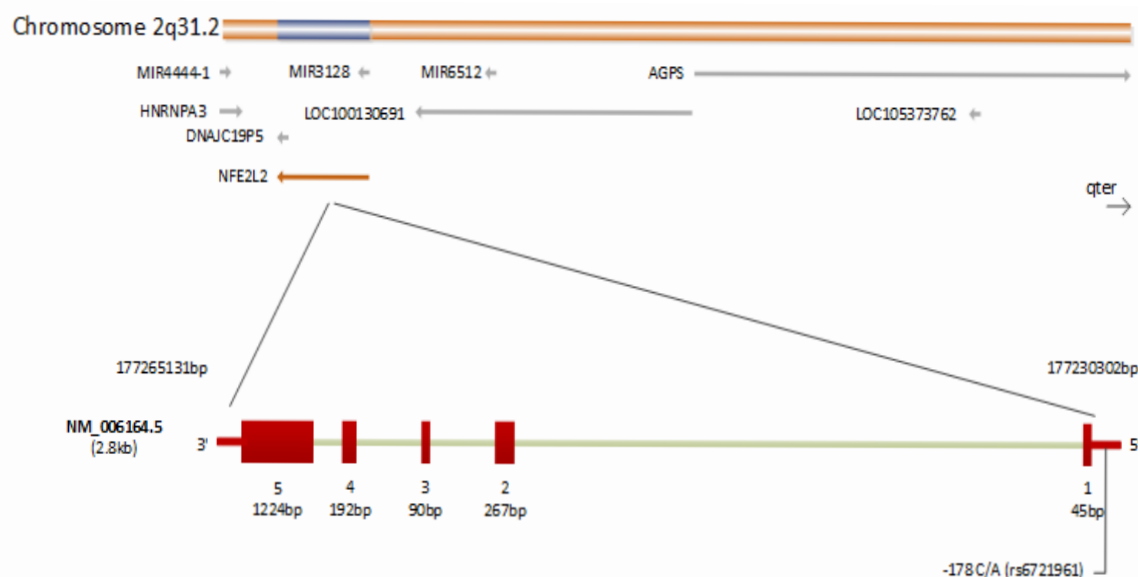


Figure 1. Nrf2 gene

research studies (**table 1**) have reported the association of Nrf2 (-617C/A) polymorphism, either with the level of oxidative stress biomarkers in plasma of ESRD patients (54), or the susceptibility towards ESRD development and prognosis (54, 56). In the study conducted on 256 Serbian

Even though aforementioned studies contributed to clearer positioning of Nrf2 polymorphisms in the overall picture of complex interactions present in ESRD, there are still numerous unexplored issues in this field. Ongoing, promising developments in therapeutic interventions are

**Table 1.** The association of SOD2, GPX1 and Nrf2 polymorphisms with CKD risk, progression and mortality

Reference	Location	Study group	SOD2 rs4880	GPX1 rs1050450	Nrf2 rs6721961
Crawford et al. 2011.	Australia	185 CKD patients	<i>Ala/Val</i> and <i>Val/Val</i> genotypes had a significantly greater eGFR decline compared to those with the <i>Ala/Ala</i> genotype	No significant association with the progression of CKD	NR
Crawford et al. 2012.	Australia	230 CKD patients / 224 controls	No significant association with CKD risk	<i>Leu/Leu</i> genotype was associated with increased risk of CKD and lower eGFR	NR
Shimoyama et al. 2014.	Japan	216 ESRD patients / 464 controls	NR	NR	No significant association with ESRD risk, overall and cardiovascular survival
Chao et al. 2016.	China	671 ESRD patients / 780 controls	<i>Ala/Ala</i> genotype was associated with increased risk of ESRD	No individual significant association with ESRD risk. GPX1 <i>Leu/Leu</i> genotype was associated with increased risk of ESRD when combined with PPAR - $\gamma$ <i>G/G</i> genotype.	NR
Abbasi et al. 2018.	Iran	280 CKD patients / 280 T2D controls	<i>Val/Val</i> genotype was associated with increased risk of CKD	NR	NR
Jerotic et al. 2019.	Serbia	256 ESRD patients / 374 controls	<i>Val/Val</i> genotype was associated with increased risk of ESRD. No significant association with overall and cardiovascular survival.	No individual significant association with ESRD risk. GPX1 <i>Leu/Leu</i> genotype was associated with increased risk of ESRD when combined with SOD2 <i>Val/Val</i> genotype. GPX1 ( <i>Leu/Leu</i> ) genotype was associated with longer cardiovascular survival.	No individual significant association with ESRD risk. Nrf2 <i>C/C</i> genotype was associated with increased risk of ESRD when combined with SOD2 <i>Val/Val</i> genotype. Nrf2 ( <i>C/C</i> ) genotype was associated with longer overall survival when combined with GPX1 ( <i>Leu/Leu</i> ) genotype.
Corredor et al. 2020.	Spain	722 CKD patients / 172 controls	No significant association with CKD risk	No significant association with CKD risk	NR

Abbreviations: Not reported (NR)

ESRD patients and 374 healthy controls, Nrf2 (-617C/A) polymorphism was not associated with the oxidative phenotype of these patients, nor with the ESRD risk (54). In the same study, Nrf2 C/C genotype had an impact by increasing risk of ESRD, in combination with SOD2 *Val/Val* genotype, and longer overall survival, after being combined with GPX1 *Leu/Leu* genotype (54). Nevertheless, no individual association of Nrf2 (-617C/A) polymorphism was observed in relation to the risk, overall and cardiovascular survival among these patients (54). These results are similar to those obtained in a study of Shimoyama et al. using Japanese cohort of 216 ESRD patients and 464 controls (56). In this study, aforementioned Nrf2 polymorphism did not have an impact on the risk of ESRD development or overall/cardiovascular survival. The only significant association was found between other Nrf2 polymorphism, G/A rs35652124 and cardiovascular survival in dialysed patients (56).

underway, with several possible candidates that might be beneficial to patients with diminished Nrf2 expression and function, as present in variant, Nrf2 A carriers. A common feature of these compounds is the potential for inducing Nrf2 pathway. One example of such a compound is bardoxolone methyl, which effects on CKD patients were examined in clinical trial investigations (59). Measurable improvement in clinical parameters of CKD patients determined in those studies included a significant increase in GFR, as well as creatinine clearance (59). These effects could be attributed to bardoxolone methyl's activation of the Nrf2 pathway, and subsequent restoration of redox imbalance (59).

## SOD2 Polymorphism and CKD

SOD and GPX are antioxidant enzymes that work in a mutually supportive fashion in scavenging superoxide



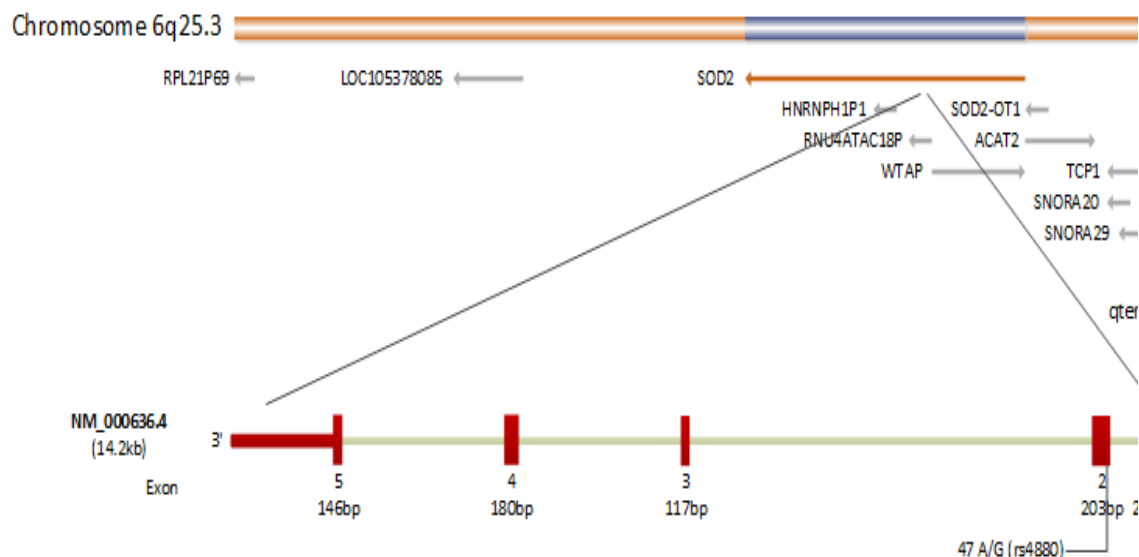


Figure 2. SOD2 gene

radicals and decreasing oxidative stress, abundantly produced in CKD. SOD converts superoxide ( $O_2^-$ ) to hydrogen peroxide ( $H_2O_2$ ) with three isoforms being described in the literature (17, 60, 61). Two SOD isoforms contain copper (*Cu*) and zinc (*Zn*) within the active site: intracellular SOD1 - present in red blood cells, and extracellular SOD3 - present in interstitium and extracellular fluids (17, 62, 63). On the other hand, SOD2 contains a manganese within the active site (MnSOD) and is positioned within mitochondria, where it scavenges superoxide radicals (22, 63). In humans, the polymorphisms have been identified in all these three SOD isoforms (64). Nevertheless, the most highly active SOD isoform in the human kidney is SOD2 isoform and its rs4880 functional polymorphism has the highest implication in human diseases investigated so far. Therefore, the main focus of this review will be the association of SOD2 rs4880 polymorphism and CKD risk and progression.

MnSOD gene is positioned on chromosome 6q25 (34). The SOD2 rs4880 SNP, located in exon 2, is characterised by a C > T nucleotide substitution resulting in the amino acid alanine (*Ala*) > valine (*Val*) substitution at position 16 (SOD2 *Ala16Val* genotype) (64, 65). Importantly, the *Val* variant of this SNP compromises the ability of SOD2 to neutralize superoxide radicals within the mitochondria by altering SOD2 expression, as well as mitochondrial transport (34, 64). In this line, there are indications that the *Ala* allele, accompanied with higher SOD2 mitochondrial activity, may provide better protection against the progression of oxidative stress associated diseases such as CKD (54, 61).

To the best of our knowledge, six original investigations have so far tested an association between SOD2 rs4880 SNP and either CKD or ESRD, with contradictory results (table 1). In the Crawford et al. study, among 185 CKD patients included, those carrying low activity SOD2, *Val* allele had a marked drop in GFR, in comparison with patients carrying the *Ala/Ala* genotype (61, 66). In addition to the influence that *Val* allele has in terms of faster

progression of CKD measured through the GFR deterioration, it has been shown that it might influence the risk of the CKD and ESRD development, as well. Namely, in a study of Jerotić et al., SOD2 *Val/Val* genotype resulted in a greater susceptibility towards ESRD (54). Similarly, in an Iranian cohort of 280 type 2 diabetic (T2D) patients with CKD and 280 T2D controls, patients with the *Val/Val* genotype exhibited higher CKD risk than those having the *Ala/Ala* or *Ala/Val* genotypes (67). Additionally, several studies consistently confirmed that the *Val* allele increases the risk and the progression of diabetic nephropathy with a faster decline in GFR (68–71). On the other hand, in a cohort of 671 ESRD patients and 780 healthy controls from China, the *Val* allele appeared to be protective, given that the *Ala/Ala* genotype correlated with increased risk of ESRD (72). This could be due to differences in the genotype distribution between Asian and Caucasian population. Namely, the frequency of the *Ala/Ala* genotype in Caucasians was 32%, whereas in the Asian population it was only 2% (54, 66, 72). On the contrary to the aforementioned reports, two studies conducted in Australia (230 CKD patients and 224 healthy controls) and Spain (722 CKD patients and 172 controls), reported no association between SOD2 rs4880 polymorphism and CKD risk (61, 73).

As discussed above, increased production of ROS is accompanied by impaired antioxidant defense in CKD. However, the functional significance of antioxidant enzyme polymorphisms in terms of the level of oxidative stress byproducts in these patients still remains elusive. Jerotić et al. have also shown that polymorphisms of enzymes front-line protection against free radicals, such as SOD2, can raise byproducts of oxidative damage in ESRD patients (54). In addition to having higher susceptibility to ESRD development, dialysis patients with the SOD2 *Val/Val* genotype also had higher plasma levels of byproducts of protein oxidative damage (thiol and carbonyl groups, advanced oxidation byproducts (AOPP, nitrotyrosine), as well as lipid oxidative damage (MDA and MDA adducts) (54). The substantial role of SOD2 polymorphisms on

oxidative stress in patients with impaired kidney function was also confirmed in a large study conducted on patients with diabetes mellitus induced nephropathy (70).

The most widely studied impact of SOD2 rs4880 polymorphism, in terms of kidney pathology, was the one

an increase in selenium (80).

The following GPX1 rs1050450 polymorphism has been extensively studied in human diseases, mostly with regard to breast, lung and bladder cancer pathology (64); however, to date, only five studies have repor-

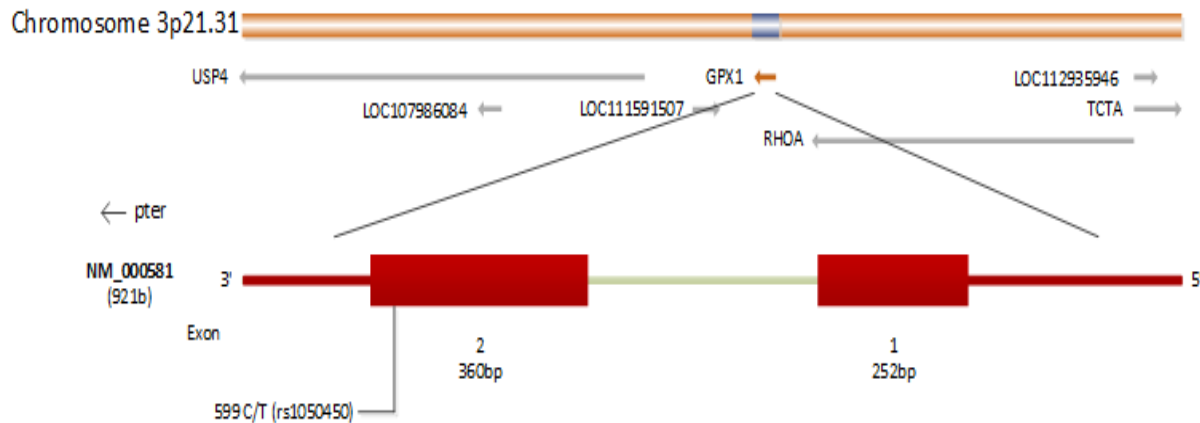


Figure 3. GPX1 gene

associated with the onset and progression of diabetic nephropathy (66, 68–71). These studies consistently showed that the low activity, *Val* allele affects the higher susceptibility to diabetic nephropathy and its progression. However, association of SOD2 polymorphism with an antioxidant status, levels of oxidative stress biomarkers and their implications to CKD triggered by either diabetes or other factors, still need to be clarified in future studies.

## GPX1 Polymorphism and CKD

Dismutation of O<sub>2</sub><sup>-</sup> to H<sub>2</sub>O<sub>2</sub> by SOD is followed by H<sub>2</sub>O<sub>2</sub> reduction to water, which is a process primarily mediated by another antioxidant enzyme, glutathione peroxidase (17). GPX is a selenoprotein containing the rare amino acid selenocysteine at its active site (17, 74). Noteworthy, the activity of GPX enzyme is significantly altered in all stages of CKD (22). Multiple studies consistently reported marked decrease in GPX activity from the CKD onset through the progression of uremia, with the lowest activity found in ESRD patients (21, 22, 75, 76). Several isozymes of glutathione peroxidase family that vary in cellular location and substrate specificity have been identified so far (17, 77). GPX1 is the most abundant isozyme in the glutathione peroxidases family (17, 35, 77). This enzyme is cytosolic and produced in all tissues (17, 64, 77). Nevertheless, the main sources of GPX1 are erythrocytes, liver and kidneys (17, 64, 77).

The GPX1 gene is positioned on the 3p21.3 chromosome and has two exons (78). Of a particular importance is the functional GPX1 rs1050450 polymorphism, which induces C > T substitution changing the amino acid proline (*Pro*) with leucine (*Leu*) at position 198 (GPX1 Pro198Leu genotype) (64). The *Leu* allele of GPX1 gene confers with significantly impaired enzyme activity, which was found *in vitro* and also confirmed in humans (35, 79). Moreover, *Leu* allele makes this enzyme less responsive to

ted on its association with CKD (**table 1**). In addition to SOD2 rs4880 polymorphism as discussed above, GPX1 rs1050450 polymorphism has also been associated with an increased risk and progression of CKD (75). According to these results, significantly higher number of CKD patients had the GPX1 *Leu/Leu* genotype compared to controls (66). Moreover, *Leu/Leu* genotype was associated with lower eGFR in CKD patients (66). On the other hand, there are several studies reporting no individual association between this polymorphism and the risk of CKD, although its influence appeared significant when combined with the other gene polymorphisms including SOD2 and PPAR- $\gamma$  (54, 72). These findings illustrate that the antioxidant system is comprised of a range of different interactions, thus eliminating the possibility of identifying a distinct variable, which could be used as a single marker of disease. To this effect, Jerotic et al. were the first research group to explore the association between GPX1 rs1050450 polymorphism and oxidative phenotype among ESRD patients (54). They found no significant impact of this polymorphism on either one of estimated biomarkers of oxidative stress in plasma of these patients (54). However, another study demonstrated correlation of other GPX1 gene polymorphisms (rs3448, rs9819758, rs8179164 and rs1987628) and oxidative stress byproducts (AOPP and isoprostanes) (54, 81). In this study, T allele of GPX1 rs3448 gene had an impact on higher susceptibility to ESRD among diabetic patients, as well as, higher levels of AOPP and isoprostanes, while other examined GPX1 polymorphisms did not show any significant association (54). The aforementioned study of Jerotic et al. is also the only one examining the potential predictive influence of GPX1 rs1050450 polymorphism on ESRD patients survival (54). Unexpectedly, low activity, GPX1 *Leu/Leu*, genotype contributed to longer overall, as well as cardiovascular survival in ESRD patients (54). Even though there are a few studies suggesting the adverse influence of GPX1 *Leu* allele on CVDs, such as coronary

heart disease, calcification of coronary arteries and increased intima-media thickness (74), there are also several studies conversely reporting that this allele may be protective (54, 74, 82, 83). Namely, it has been shown that *Leu* allele may reduce the risk of thoracic aortic aneurysm in patients with hypertension (74, 83). Moreover, Soerensen et al. showed that GPX1 *Leu* allele influenced longer survival in the oldest population (82). Overall, there is a subtle balance between the beneficial and deleterious effects of ROS, with complex signalling mechanisms that need to be explored further, in order to provide better understanding and enable clinical translation.

## Conclusion

Already recognized causes of CKD converge at the final common pathway – oxidative stress mediated loss of renal function. Therefore, individual susceptibility to CKD seems closely linked to functional variations of genes, encoding antioxidant regulatory and catalytic proteins. The results reported so far in the literature suggest that these three different polymorphisms could be important targets for better patient outcomes. Identifying important polymorphism in CKD patients could lead to better stratification and more personalised treatment of CKD patients. Given that CKD is a multifactorial disease, a number of different biomarkers based on the presence of polymorphisms and biomarkers of oxidative stress should be included as part of the risk stratification strategies. The multimarker panel could be explored for prediction of CKD patients' survival, or as an indicator of the need for antioxidant therapy on individual patient basis. New promising therapeutic possibilities are underway that might be beneficial to CKD patients with diminished Nrf2, SOD2 or GPX1 expression and function.

## Literature

1. Levin A, Stevens PE, Bilous RW, Coresh J, De Francisco ALM, De Jong PE, et al. Kidney Disease: Improving Global Outcomes (KDIGO) CKD Work Group. KDIGO 2012 clinical practice guideline for the evaluation and management of chronic kidney disease. *Kidney Int Suppl.* 2013; 3(1):1–150.
2. Vajdic CM, McDonald SP, McCredie MRE, Van Leeuwen MT, Stewart JH, Law M, et al. Cancer incidence before and after kidney transplantation. *Jama.* 2006; 296(23):2823–31.
3. Tonelli M, Wiebe N, Culleton B, House A, Rabbat C, Fok M, et al. Chronic kidney disease and mortality risk: a systematic review. *J Am Soc Nephrol.* 2006; 17(7):2034–47.
4. Thompson S, James M, Wiebe N, Hemmelgarn B, Manns B, Klarenbach S, et al. Cause of death in patients with reduced kidney function. *J Am Soc Nephrol.* 2015; 26(10):2504–11.
5. Fiorillo C, Oliviero C, Rizzuti G, Nediani C, Pacini A, Nassi P. Oxidative stress and antioxidant defenses in renal patients receiving regular haemodialysis. *Clin Chem Lab Med.* 1998; 36(3):149–53.
6. Vaziri ND. Oxidative stress in uremia: nature, mechanisms, and potential consequences. In: *Seminars in nephrology.* Elsevier; 2004. p. 469–73.
7. Himmelfarb J, Stenvinkel P, Ikizler TA, Hakim RM. The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int.* 2002; 62(5):1524–38.
8. Oberg BP, Mcmenamin E, Lucas FL, Mcmonagle E, Morrow J, Ikizler TA, et al. Increased prevalence of oxidant stress and inflammation in patients with moderate to severe chronic kidney disease. *Kidney Int.* 2004; 65(3):1009–16.
9. Vaziri ND, Oveisi F, Ding Y. Role of increased oxygen free radical activity in the pathogenesis of uremic hypertension. *Kidney Int.* 1998; 53(6):1748–54.
10. Sies H. Oxidative stress: from basic research to clinical application. *Am J Med.* 1991; 91(3):S31–8.
11. Sies H. Oxidative stress: eustress and distress in redox homeostasis. In: *Stress: physiology, biochemistry, and pathology.* Elsevier; 2019. p. 153–63.
12. Massy ZA, Ceballos I, Chadefaux-Vekemens B, Nguyen-Khoa T, Descamps-Latscha B, Drüeke TB, et al. Homocyst (e) ine, oxidative stress, and endothelium function in uremic patients. *Kidney Int.* 2001; 59(S78):S243.
13. Malhotra JD, Kaufman RJ. Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? *Antioxid Redox Signal.* 2007; 9(12):2277–94.
14. Chin MP, Reisman SA, Bakris GL, O'grady M, Linde PG, McCullough PA, et al. Mechanisms contributing to adverse cardiovascular events in patients with type 2 diabetes mellitus and stage 4 chronic kidney disease treated with bardoxolone methyl. *Am J Nephrol.* 2014; 39(6):499–508.
15. Ward RA, McLeish KR. Polymorphonuclear leukocyte oxidative burst is enhanced in patients with chronic renal insufficiency. *J Am Soc Nephrol.* 1995; 5(9):1697–702.
16. Galvan DL, Green NH, Danesh FR. The hallmarks of mitochondrial dysfunction in chronic kidney disease. *Kidney Int.* 2017; 92(5):1051–7.
17. Laher I. *Systems biology of free radicals and antioxidants.* Springer; 2014.
18. Nguyen AT, Lethias C, Zingraff J, Herbelin A, Naret C, Descamps-Latscha B. Hemodialysis membrane-induced activation of phagocyte oxidative metabolism detected in vivo and in vitro within microamounts of whole blood. *Kidney Int.* 1985; 28(2):158–67.
19. Tbahriti HF, Kaddous A, Bouchenak M, Mekki K. Effect of different stages of chronic kidney disease and renal replacement therapies on oxidant-antioxidant balance in uremic patients. *Biochem Res Int.* 2013; 2013.
20. Girelli D, Olivieri O, Stanzial AM, Azzini M, Lupo A, Bernich P, et al. Low platelet glutathione peroxidase activity and serum selenium concentration in patients with chronic renal failure: relations to dialysis treatments, diet and cardiovascular complications. *Clin Sci.* 1993; 84(6):611–7.
21. Ceballos-Picot I, Witko-Sarsat V, Merad-Boudia M, Nguyen AT, Thévenin M, Jaudon MC, et al. Glutathione antioxidant system as a marker of oxidative stress in chronic renal failure. *Free Radic Biol Med.* 1996; 21(6):845–53.
22. Sung C-C, Hsu Y-C, Chen C-C, Lin Y-F, Wu C-C. Oxidative stress and nucleic acid oxidation in patients with chronic kidney disease. *Oxid Med Cell Longev.* 2013; 2013.
23. Daenen K, Andries A, Mekahli D, Van Schepdael A, Jouret F, Bammens B. Oxidative stress in chronic kidney disease. *Pediatr Nephrol.* 2019; 34(6):975–91.
24. Ward RA, Ouseph R, Mcleish KR. Effects of high-flux hemodialysis on oxidant stress. *Kidney Int.* 2003; 63(1):353–9.
25. Jankowska M, Rutkowski B, Dębska-Ślizień A. Vitamins and microelement bioavailability in different stages of chronic kidney disease. *Nutrients.* 2017; 9(3):282.
26. Davies KJ. Protein damage and degradation by oxygen radicals. I. general aspects. *J Biol Chem.* 1987; 262(20):9895–901.
27. Wolff SP, Garner A, Dean RT. Free radicals, lipids and protein degradation. *Trends Biochem Sci.* 1986; 11(1):27–31.
28. Imlay JA, Linn S. DNA damage and oxygen radical toxicity. *Science (80- ).* 1988; 240(4857):1302–9.
29. Mimić-Oka J, Savić-Radojević A, Plješa-Ercegovac M, Opačić M, Simić T, Dimković N, et al. Evaluation of oxidative stress after repeated intravenous iron supplementation. *Ren Fail.* 2005; 27(3):345–51.
30. Choi B, Kang K-S, Kwak M-K. Effect of redox modulating NRF2 activators on chronic kidney disease. *Molecules.* 2014;



- 19(8):12727–59.
31. Tucker PS, Dalbo VJ, Han T, Kingsley MI. Clinical and research markers of oxidative stress in chronic kidney disease. *Biomarkers*. 2013; 18(2):103–15.
  32. Grieve DJ, Pljesa-Ercegovac M, Savic-Radojevic A, Damjanovic T, Dimkovic N, McClements L, et al. Research Article GSTM1 Modulates Expression of Endothelial Adhesion Molecules in Uremic Milieu. 2021;
  33. Marzec JM, Christie JD, Reddy SP, Jedlicka AE, Vuong H, Lanken PN, et al. Functional polymorphisms in the transcription factor NRF2 in humans increase the risk of acute lung injury. *FASEB J*. 2007; 21(9):2237–46.
  34. Sutton A, Khoury H, Prip-Buus C, Capanec C, Pessayre D, Degoul F. The Ala16Val genetic dimorphism modulates the import of human manganese superoxide dismutase into rat liver mitochondria. *Pharmacogenet Genomics*. 2003; 13(3):145–57.
  35. Zheikova T V, Golubenko M V, Buikin S V, Botkina OY, Makeeva OA, Lezhnev AA, et al. Glutathione peroxidase 1 (GPX1) single nucleotide polymorphism Pro198→ Leu: Association with life span and coronary artery disease. *Mol Biol*. 2012; 46(3):433–7.
  36. Hamanishi T, Furuta H, Kato H, Doi A, Tamai M, Shimomura H, et al. Functional variants in the glutathione peroxidase-1 (GPx-1) gene are associated with increased intima-media thickness of carotid arteries and risk of macrovascular diseases in Japanese type 2 diabetic patients. *Diabetes*. 2004; 53(9):2455–60.
  37. Li W, Khor TO, Xu C, Shen G, Jeong W-S, Yu S, et al. Activation of Nrf2-antioxidant signaling attenuates NFκB-inflammatory response and elicits apoptosis. *Biochem Pharmacol*. 2008; 76(11):1485–9.
  38. Tonelli C, Chio IIC, Tuveson DA. Transcriptional regulation by Nrf2. *Antioxid Redox Signal*. 2018; 29(17):1727–45.
  39. Kobayashi M, Yamamoto M. Nrf2-Keap1 regulation of cellular defense mechanisms against electrophiles and reactive oxygen species. *Adv Enzyme Regul*. 2006; 46:113–40.
  40. Ihoriya C, Satoh M, Komai N, Sasaki T, Kashihara N. Nuclear factor erythroid 2-related factor 2 is activated by rosuvastatin via p21cip1 upregulation in endothelial cells. *Biochem Pharmacol (Los Angel)*. 2014; 4(157):501–2167.
  41. Stockler-Pinto MB, Fouques D, Soulage CO, Croze M, Mafra D. Indoxyl sulfate and p-cresyl sulfate in chronic kidney disease. Could these toxins modulate the antioxidant Nrf2-Keap1 pathway? *J Ren Nutr*. 2014; 24(5):286–91.
  42. Yamamoto T, Yoh K, Kobayashi A, Ishii Y, Kure S, Koyama A, et al. Identification of polymorphisms in the promoter region of the human NRF2 gene. *Biochem Biophys Res Commun*. 2004; 321(1):72–9.
  43. Nishinaka T, Ichijo Y, Ito M, Kimura M, Katsuyama M, Iwata K, et al. Curcumin activates human glutathione S-transferase P1 expression through antioxidant response element. *Toxicol Lett*. 2007; 170(3):238–47.
  44. Zhang DD, Lo S-C, Sun Z, Habib GM, Lieberman MW, Hannink M. Ubiquitination of Keap1, a BTB-Kelch substrate adaptor protein for Cul3, targets Keap1 for degradation by a proteasome-independent pathway. *J Biol Chem*. 2005; 280(34):30091–9.
  45. Itoh K, Wakabayashi N, Katoh Y, Ishii T, Igarashi K, Engel JD, et al. Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. *Genes Dev*. 1999; 13(1):76–86.
  46. Kim HJ, Vaziri ND. Contribution of impaired Nrf2-Keap1 pathway to oxidative stress and inflammation in chronic renal failure. *Am J Physiol Physiol*. 2010; 298(3):F662–71.
  47. Surh Y-J, Kundu JK, Na H-K. Nrf2 as a master redox switch in turning on the cellular signaling involved in the induction of cytoprotective genes by some chemopreventive phytochemicals. *Planta Med*. 2008; 74(13):1526–39.
  48. Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol*. 2007; 47:89–116.
  49. Yoh K, Hirayama A, Ishizaki K, Yamada A, Takeuchi M, Yamagishi S, et al. Hyperglycemia induces oxidative and nitrosative stress and increases renal functional impairment in Nrf2 deficient mice. *Genes to Cells*. 2008;13(11):1159–70.
  50. Jiang T, Huang Z, Lin Y, Zhang Z, Fang D, Zhang DD. The protective role of Nrf2 in streptozotocin-induced diabetic nephropathy. *Diabetes*. 2010; 59(4):850–60.
  51. Jiang T, Tian F, Zheng H, Whitman SA, Lin Y, Zhang Z, et al. Nrf2 suppresses lupus nephritis through inhibition of oxidative injury and the NF-κB-mediated inflammatory response. *Kidney Int*. 2014; 85(2):333–43.
  52. Yoh K, Itoh K, Enomoto A, Hirayama A, Yamaguchi N, Kobayashi M, et al. Nrf2-deficient female mice develop lupus-like autoimmune nephritis. *Kidney Int*. 2001; 60(4):1343–53.
  53. Liu M, Grigoryev DN, Crow MT, Haas M, Yamamoto M, Reddy SP, et al. Transcription factor Nrf2 is protective during ischemic and nephrotoxic acute kidney injury in mice. *Kidney Int*. 2009; 76(3):277–85.
  54. Jerotic D, Matic M, Suvakov S, Vucicevic K, Damjanovic T, Savic-Radojevic A, et al. Association of Nrf2, SOD2 and GPX1 Polymorphisms with Biomarkers of Oxidative Distress and Survival in End-Stage Renal Disease Patients. *Toxins (Basel)*. 2019; 11(7):431.
  55. Cho H-Y, Marzec J, Kleeberger SR. Functional polymorphisms in Nrf2: implications for human disease. *Free Radic Biol Med*. 2015; 88:362–72.
  56. Shimoyama Y, Mitsuda Y, Tsuruta Y, Hamajima N, Niwa T. Polymorphism of Nrf2, an antioxidative gene, is associated with blood pressure and cardiovascular mortality in hemodialysis patients. *Int J Med Sci*. 2014; 11(7):726.
  57. Cho H-Y. Genomic structure and variation of nuclear factor (erythroid-derived 2)-like 2. *Oxid Med Cell Longev*. 2013;2013.
  58. Marczak ED, Marzec J, Zeldin DC, Kleeberger SR, Brown NJ, Pretorius M, et al. Polymorphisms in the transcription factor NRF2 and forearm vasodilator responses in humans. *Pharmacogenet Genomics*. 2012; 22(8):620.
  59. Pergola PE, Raskin P, Toto RD, Meyer CJ, Huff JW, Grossman EB, et al. Bardoxolone methyl and kidney function in CKD with type 2 diabetes. *N Engl J Med*. 2011; 365(4):327–36.
  60. Zelko IN, Mariani TJ, Folz RJ. Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. *Free Radic Biol Med*. 2002; 33(3):337–49.
  61. Crawford A, Fassett RG, Coombes JS, Kunde DA, Ahuja KDK, Robertson IK, et al. Glutathione peroxidase, superoxide dismutase and catalase genotypes and activities and the progression of chronic kidney disease. *Nephrol Dial Transplant*. 2011; 26(9):2806–13.
  62. Nozik-Grayck E, Suliman HB, Piantadosi CA. Extracellular superoxide dismutase. *Int J Biochem Cell Biol*. 2005; 37(12):2466–71.
  63. Azadmanesh J, Borgstahl GEO. A review of the catalytic mechanism of human manganese superoxide dismutase. *Antioxidants*. 2018; 7(2):25.
  64. Crawford A, Fassett RG, Geraghty DP, Kunde DA, Ball MJ, Robertson IK, et al. Relationships between single nucleotide polymorphisms of antioxidant enzymes and disease. *Gene*. 2012; 501(2):89–103.
  65. Rosenblum JS, Gilula NB, Lerner RA. On signal sequence polymorphisms and diseases of distribution. *Proc Natl Acad Sci*. 1996; 93(9):4471–3.
  66. Crawford A. Influence of antioxidant genotype and antioxidant status on progression of chronic kidney disease. University of Tasmania; 2010.
  67. Abbasi M, Daneshpour MS, Hedayati M, Mottaghi A, Pourvali K, Azizi F. The relationship between MnSOD Val16Ala gene polymorphism and the level of serum total antioxidant capacity with the risk of chronic kidney disease in type 2 diabetic patients: a nested case-control study in the Tehran lipid glucose study. *Nutr Metab (Lond)*. 2018; 15(1):25.
  68. Möllsten A, Marklund SL, Wessman M, Svensson M, Forsblom C, Parkkonen M, et al. A functional polymorphism in the manganese superoxide dismutase gene and diabetic nephropathy. *Diabetes*. 2007; 56(1):265–9.
  69. Möllsten A, Jorsal A, Lajer M, Vionnet N, Tarnow L. The V16A



- polymorphism in SOD2 is associated with increased risk of diabetic nephropathy and cardiovascular disease in type 1 diabetes. *Diabetologia*. 2009; 52(12):2590–3.
70. Mohammedi K, Bellili-Muñoz N, Driss F, Roussel R, Seta N, Fumeron F, et al. Manganese superoxide dismutase (SOD2) polymorphisms, plasma advanced oxidation protein products (AOPP) concentration and risk of kidney complications in subjects with type 1 diabetes. *PLoS One*. 2014; 9(5):e96916.
  71. Nomiya T, Tanaka Y, Piao L, Nagasaka K, Sakai K, Ogihara T, et al. The polymorphism of manganese superoxide dismutase is associated with diabetic nephropathy in Japanese type 2 diabetic patients. *J Hum Genet*. 2003; 48(3):138.
  72. Chao C-T, Chen Y-C, Chiang C-K, Huang J-W, Fang C-C, Chang C-C, et al. Interplay between superoxide dismutase, glutathione peroxidase, and peroxisome proliferator activated receptor gamma polymorphisms on the risk of end-stage renal disease among Han Chinese patients. *Oxid Med Cell Longev*. 2016; 2016.
  73. Corredor Z, da Silva Filho MI, Rodríguez-Ribera L, Velázquez A, Hernández A, Catalano C, et al. Genetic Variants Associated with Chronic Kidney Disease in a Spanish Population. *Sci Rep*. 2020; 10(1):1–11.
  74. Lubos E, Loscalzo J, Handy DE. Glutathione peroxidase-1 in health and disease: from molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal*. 2011; 15(7):1957–97.
  75. Crawford A, Fassett RG, Coombes JS, Kunde DA, Ahuja KDK, Robertson IK, et al. Relationship between antioxidant enzyme genotype and activity and kidney function: a case-control study. *Clin Nephrol*. 2012; 78(2):135–44.
  76. Mimić-Oka J, Simić T, Djukanović L, Reljić Z, Davicević Z. Alteration in plasma antioxidant capacity in various degrees of chronic renal failure. *Clin Nephrol*. 1999; 51(4):233–41.
  77. Zotova E V, Savost'ianov K V, Chistiakov DA, Bursa TR, Galeev I V, Stokov IA, et al. Search for the association of polymorphic markers for genes coding for antioxidant defense enzymes, with development of diabetic polyneuropathies in patients with type 1 diabetes mellitus. *Mol Biol (Mosk)*. 2004; 38(2):244–9.
  78. Tang N-P, Wang L-S, Yang L, Gu H-J, Sun Q-M, Cong R-H, et al. Genetic variant in glutathione peroxidase 1 gene is associated with an increased risk of coronary artery disease in a Chinese population. *Clin Chim acta*. 2008; 395(1–2):89–93.
  79. Elelaimy IA, Shehata EL, Abdel-Hamid MA. Study the association between glutathione peroxidase-1 gene in patients with hepatocellular carcinoma in Egypt. *J Biosci Appl Res*. 2016; 2(6):346–51.
  80. Hu YJ, Diamond AM. Role of glutathione peroxidase 1 in breast cancer: loss of heterozygosity and allelic differences in the response to selenium. *Cancer Res*. 2003; 63(12):3347–51.
  81. Mohammedi K, Patente TA, Bellili-Muñoz N, Driss F, Le Nagard H, Fumeron F, et al. Glutathione peroxidase-1 gene (GPX1) variants, oxidative stress and risk of kidney complications in people with type 1 diabetes. *Metabolism*. 2016; 65(2):12–9.
  82. Soerensen M, Christensen K, Stevnsner T, Christiansen L. The Mn-superoxide dismutase single nucleotide polymorphism rs4880 and the glutathione peroxidase 1 single nucleotide polymorphism rs1050450 are associated with aging and longevity in the oldest old. *Mech Ageing Dev*. 2009; 130(5):308–14.
  83. Kato K, Oguri M, Kato N, Hibino T, Yajima K, Yoshida T, et al. Assessment of genetic risk factors for thoracic aortic aneurysm in hypertensive patients. *Am J Hypertens*. 2008; 21(9):1023–7.